

# A long way: History of the prophylactic papillomavirus vaccine

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## 1. History of papillomaviruses as infectious agents in animals and humans

For more than one and a half centuries, papillomaviruses have been recognized as causative agents for benign and malignant disease in humans as well as in a number of animal species [43]. It took more than five decades to provide sufficient evidence for the general acceptance of HPV as necessary cause for the development of anogenital cancer in humans. At the end of this long road of extensive studies on papillomaviruses by countless scientists worldwide there is finally a prophylactic papillomavirus vaccine at hand. Clearly, the scientific progress in vaccine development was hampered by the fact that papillomaviruses cannot be readily cultivated *in vitro*. In this review we will discuss the path that led to the introduction of the prophylactic vaccine to the public.

Earliest reports about a putative infectious agent being involved in the development of cervical cancers came from observations made by the Italian physician Domenico Rigoni-Stern [51]. He noted that cancer of the womb is found most frequently among women in their fourth and fifth decade and he described behavioural risk factors for this cancer that are still being in the focus of modern epidemiology: age at first sexual intercourse and promiscuity. Despite an imprecise baseline of his study (Rigoni-Stern did not differentiate between cancer of the endometrial and cervical cancer) the link to cervical cancer was later confirmed by other

investigators in the 1950s eventually linking cervical cancer to sexually transmitted agents [22,62,63]. In the following, a number of known pathogens were suspected to play a role in cervical cancer development. More refined investigations were subsequently able to rule out an involvement of *Treponema pallidum* (Syphilis) or Herpes Simplex Virus 2 and others in anogenital cancer [66].

Another line of evidence for papillomaviruses being responsible for tumor development was established by experimental transmission of warts and condylomas in man. As early as in the 1890s several investigators induced skin warts in volunteers by the inoculation of hand wart extracts [41,64]. In 1907 Ciuffo demonstrated that a filterable agent is able to induce hand warts [10]. Similar experiments were later performed for condylomas by Waelsch [67]. With the introduction of electron microscopy, virus particles were identified in extracts of hand warts and condylomas. Already in the early 1930s Shope described a virus inducing papillomas in cottontail and domestic rabbits [60]. Shope observed that experimentally induced papillomas in domestic rabbits progress to cancer in high frequency providing the first example of a DNA virus as causative agent for cancers in mammals. Later, Ito and Evans demonstrated that papillomas and tumors can be induced solely with naked viral DNA [33].

In 1976 zur Hausen noted that the epidemiological patterns of cervical cancer and condylomas showed striking similarities [72]. As papillomaviruses had been already tracked down for causing condylomas it was postulated that they might also be causatively involved in the development of cervical cancer. With the onset of molecular biology, a number of genital papillo-

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mavirus types were isolated and cloned into plasmid vectors in the following. HPV 6 DNA was cloned from condylomas, the closely related HPV 11 was isolated from laryngeal papillomas but also detected in genital warts [23,24,59]. With these low-risk papillomavirus genomes at hand, it was possible to prove, for the first time, presence of papillomavirus-related DNA in biopsies of cervical cancer. Shortly after, the two most prevalent high-risk HPV types 16 and 18 were cloned from tumor biopsies [4,15].

## 2. History of PV vaccination

### 2.1. Therapeutic approaches

Over the centuries, there have been numerous and sometimes venturesome attempts to treat skin and genital warts which makes it difficult to pinpoint the first therapeutic vaccination against PV. Early observations were made in the course of experimental induction of warts in rabbits and humans. It was noted, that warts develop only in a proportion of the individuals which was interpreted either as inefficiency of the virus to establish infections or 'resistance' of a given individual [26]. Further, it was clear that warts in animals and humans spontaneously regress. During a series of self-inoculations with wart extracts Findlay stated that he became 'immune' to wart induction [21]. It was also observed that local inflammation precedes regression of warts and that warts frequently disappear simultaneously [45]. In the 1930s and 1940s a number of therapeutic vaccination trials were performed based on the injection of autologous and heterologous wart extracts. During his trials, Biberstein inoculated hundreds of patients with filtered extracts from warts and condylomas and was able to record wart regression in the majority of the cases [3]. The likely involvement of the immune system in controlling papillomavirus infections was also underlined by the observation that genital warts but also genital cancer is more frequently found in immuno-suppressed patients [49].

The rabbit papillomavirus model proved to be particularly valuable for the study of immune therapy of PV infections. Early studies showed that regression of papillomas and transplanted CRPV-induced tumors can be induced by injection of wart extracts linking papillomas and malignant tumors to the same antigens [18]. In later years, viral antigens responsible for wart regression were identified. The early proteins E1, E2, E6, and E7 protect against virus challenge and induce regression of

already existing warts and malignant tumors [28,29]. In recent years a number of efforts have been undertaken to develop therapeutic papillomavirus vaccines some of which have entered clinical trials. It would go beyond the scope of this review to summarize all therapeutic papillomavirus vaccines, most of the approaches are directed to induce cellular immune responses against the E7 tumor antigen (for review see [14,16,61]). Synthetic E7-derived peptides have been analyzed for the treatment of low grade CIN lesion. Anti-E7 immune responses have also been induced by the use of recombinant vaccinia viruses. Interestingly, it was shown that Virus-like particles, the bases for current prophylactic vaccines, are also able to mount strong cellular immune responses likely, because they can directly interact with dendritic, antigen-presenting cells [55,57].

### 2.2. Prophylactic vaccination

Preventive vaccines are based on the induction of virus-neutralizing antibodies. In case of papillomaviruses this has first been achieved by immunization with formalin-fixed wart extracts and subsequently by inactivated purified viruses [34]. Lacking an appropriate infection model, it was difficult to determine the neutralizing activity of serum antibodies. The first *in vitro* assay for neutralization was the inhibition of focus formation of mouse cells in culture that was, for a long time, limited to BPV virions [53]. Inhibition of hemagglutination, which is based on binding of PV virions to mouse erythrocytes in a receptor-dependent manner, can be achieved by neutralizing antibodies hence this assay is considered a surrogate for measuring inhibition of infectivity [54]. A more complex neutralization assay was introduced by Kreider and coworkers: grafts of normal human tissue infected *in vitro* with HPV 11 or HPV 16 were implanted under the renal capsule of immuno-deficient mice where typical lesions develop [38]. Infection is then monitored by RT-PCR. Only very recently, a potentially high throughput neutralization assay became available which allows detection of neutralizing antibodies in an *in vitro* setting [48].

Neutralizing antibodies are directed against the major structural protein L1 and, to a much lesser degree, also against the minor capsid protein L2. In fact, in an early study *E. coli*-derived L1 protein was used for prophylactic vaccination [50] but was, despite initial promising results not followed up. The reason for the moderate success is that, for induction of neutralizing antibodies, the L1 protein is required to be presented in a correctly folded conformation. This became only

possible with the production of L1 in form of virus-like particles. In 1986 it had already been shown that purified VP1 of mouse polyomavirus spontaneously assembles into virus-like particles [56]. This inspired in the early 1990s the production of papillomavirus-like particles by different researchers. Zhou and colleagues used a vaccinia virus expression system to produce HPV 16 L1 VLPs, however the yield of this system was extremely low and did not allow further characterization of the antigen e.g. by immunological studies [70].

Shortly after, it was demonstrated that HPV 1, BPV 1 and HPV 11 VLPs can be produced much more efficiently compared to HPV 16 VLPs leading to the speculation whether this might be due to intrinsic properties of the L1 protein of different PV types [27,54,69]. After a report by Kirnbauer and colleagues it became clear, however, that the HPV 16 L1 gene of the original isolate by Dürst et al. harboured a point mutation rendering its encoded L1 protein virtually assembly defective [36]. The HPV 16 prototype genome, used by numerous researchers worldwide was derived from a tumor biopsy, in which the HPV 16 genome was integrated into the cellular genome and thus had been without any selection pressure for the maintenance of intact structural genes. When HPV 16 genomes obtained from early, virus-producing lesions were analyzed, the respective L1 proteins assembled with much greater efficiency into VLPs. Sequence analysis revealed that this difference was based on a single histidine to aspartic acid exchange at position 202. VLPs were subsequently produced successfully in insect cells, later also in yeast, in plants and even in cell free systems. Meanwhile, VLPs of several animal and human papillomaviruses have been produced, i.e. cottontail rabbit papillomavirus, rabbit oral papillomavirus, BPV 1 and 4, canine oral papillomavirus and HPV types 1, 2, 6, 8, 11, 13, 16, 18, 31, 33, 39, 45, 58 and 59 [5,9,11–13], for review see [58]. The production of VLPs in larger quantities made structural and immunological studies possible.

The compelling data obtained from several animal models fostered the concept of prophylactic vaccines against HPV infections. Virus-like particles proved to be prime candidates for prophylactic vaccination since they induce high-titer conformational antibodies both in non-human primates and in man [2,8,17,19,20,31,35,42,47]. Such antibodies proved to have neutralizing capacity [53]. By passive transfer of IgG from immunized animals it was shown that they confer protection against experimental challenge [7].

The first clinical trial with VLPs involving human subjects was initiated in 1996 (for reference see [32]).

HPV 11 VLPs were produced in insect cells and at that time laboriously purified by ultracentrifugation, a procedure that is inadequate for large scale production of a human vaccine. Nevertheless, the success of this trial spiked further HPV vaccine development. In several phase I clinical trials safety and immunogenicity of HPV 11, 16 and 18-specific VLPs generated by expression of the L1 protein in yeast or in insect cells infected with recombinant baculovirus were demonstrated [2,17,19,20,31]. Three VLP-based vaccines (HPV 16, HPV 16 + 18, HPV 6 + 11 + 16 + 18) were already brought into clinical efficacy trials. It should be noted that the vaccine provided by Merck & Co (Gardasil<sup>TM</sup>) contains in addition to the most relevant cancer-associated HPV types 16 and 18 also VLPs of the low-risk HPV types 6 and 11 aiming for protection against genital warts whereas the product developed by GlaxoSmithKline targets only the two high-risk HPVs. The data obtained from immunization of about 25,000 young women receiving either vaccine or placebo demonstrated after 4 years of follow-up protection against incident and persistent infection with HPV 6/11, and/or HPV 16 and 18 and the lesions associated therewith (external genital lesions, high grade CIN, adenocarcinoma in situ) [1,30,37,44,65]. It is as yet unclear to what extent the protection is HPV type-specific. Obviously it would be beneficial for the overall efficacy if the reactivity is not only restricted to HPV types included in the vaccine. Longer follow-up and large scale population-based efficacy trials will be required to evaluate the potential of these vaccines to reduce the incidence of cervical cancer [6,40]. Based on the existing data, however, the regulatory agencies in different countries have approved the first vaccine, the second product is expected to be licensed soon. It will be more difficult to evaluate the efficacy towards genital warts since the information about the incidence of this disease is rather scarce ([http://wrongdiagnosis.com/g/genital\\_warts](http://wrongdiagnosis.com/g/genital_warts)).

The question of acceptance of HPV-specific vaccines within different cultures also needs to be addressed [39,71]. This includes the issue whether the vaccine will be marketed as preventive measures against a sexually transmitted disease or as anti-cancer prophylaxis [46]. Other issues are the duration of immune protection and the participation of vaccinated women in Pap-screening programs HPV 16 and 18 account for about 70% of cervical cancer cases [68] hence the risk for this disease by other high-risk HPV types is far from negligible. Therefore, vaccinated women need to be educated about the necessity to keep up with the screening

program although the intervals of the visits can possibly be extended [25]. Obviously prophylactic vaccination will be highly desirable in areas where screening programs are not offered to the general population and where cervical cancer is a major public health problem. The high price clearly is prohibitive and joint efforts of scientists (developing more cost effective protocols), providers and politicians (discussing multi-tiered pricing) and public and philanthropic foundations (providing large amounts of money) are needed to make the vaccines available to the populations of highest need.

## References

- [1] K.A. Ault, Prophylactic use of quadrivalent human papillomavirus (HPV) (types 6, 11, 16, 18) L1 virus-like particle (VLP) vaccine reduces cervical intraepithelial neoplasia (CIN) 2/3 and adenocarcinoma in situ (AIS) risk, *European Journal of Cancer Supplements* **3**(4) (2005), 11.
- [2] K.A. Ault, A.R. Giuliano, R.P. Edwards, G. Tamms, L.L. Kim, J.F. Smith, K.U. Jansen, M. Allende, F.J. Taddeo, D. Skulsky and E. Barr, A phase I study to evaluate a human papillomavirus (HPV) type 18 L1 VLP vaccine, *Vaccine* **22**(23–24) (2004), 3004–3007.
- [3] Biberstein, Immunization therapy of warts, *Arch Dermatol Syphilol* **50** (1944), 12–22.
- [4] M. Boshart, L. Gissmann, H. Ikenberg, A. Kleinheinz, W. Scheurlen and H. zur Hausen, A new type of papillomavirus DNA, its presence in genital cancer biopsies and in cell lines derived from cervical cancer, *Embo J* **3**(5) (1984), 1151–1157.
- [5] J.N. Bouwes Bavincck, S. Stark, A.K. Petridis, M.E. Marugg, J. Ter Schegget, R.G. Westendorp, P.G. Fuchs, B.J. Vermeer and H. Pfister, The presence of antibodies against virus-like particles of epidermodysplasia verruciformis-associated human papillomavirus type 8 in patients with actinic keratoses, *Br J Dermatol* **142**(1) (2000), 103–109.
- [6] M.C. Bratti, A.C. Rodriguez, M. Schiffman, A. Hildesheim, J. Morales, M. Alfaro, D. Guillen, M. Hutchinson, M.E. Sherman, C. Eklund, J. Schussler, J. Buckland, L.A. Morera, F. Cardenas, M. Barrantes, E. Perez, T.J. Cox, R.D. Burk and R. Herrero, Description of a seven-year prospective study of human papillomavirus infection and cervical neoplasia among 10000 women in Guanacaste, Costa Rica, *Rev Panam Salud Publica* **15**(2) (2004), 75–89.
- [7] F. Breitburd, R. Kirnbauer, N.L. Hubbert, B. Nonnenmacher, C. Trin-Dinh-Desmarquet, G. Orth, J.T. Schiller and D.R. Lowy, Immunization with viruslike particles from cottontail rabbit papillomavirus (CRPV) can protect against experimental CRPV infection, *J Virol* **69**(6) (1995), 3959–3963.
- [8] D.R. Brown, J.T. Bryan, J.M. Schroeder, T.S. Robinson, K.H. Fife, C.M. Wheeler, E. Barr, P.R. Smith, L. Chiacchierini, A. DiCello and K.U. Jansen, Neutralization of human papillomavirus type 11 (HPV-11) by serum from women vaccinated with yeast-derived HPV-11 L1 virus-like particles: correlation with competitive radioimmunoassay titer, *J Infect Dis* **184**(9) (2001), 1183–1186.
- [9] N.D. Christensen, N.M. Cladel, C.A. Reed and R. Han, Rabbit oral papillomavirus complete genome sequence and immunity following genital infection, *Virology* **269**(2) (2000), 451–461.
- [10] Ciuffo, Innesso positivo con filtrato di verruca volgare, *Giorn Ital Mal Venereol* **48** (1907), 12–17.
- [11] A.L. Combata, M.M. Bravo, A. Touze, O. Orozco and P. Couraget, Serologic response to human oncogenic papillomavirus types 16, 18, 31, 33, 39, 58 and 59 virus-like particles in colombian women with invasive cervical cancer, *Int J Cancer* **97**(6) (2002), 796–803.
- [12] V. Cuberos, J. Perez, C.J. Lopez, F. Castro, L.V. Gonzalez, L.A. Correa, G. Sanclemente, A. Gaviria, M. Muller and G.I. Sanchez, Molecular and serological evidence of the epidemiological association of HPV 13 with focal epithelial hyperplasia: a case-control study, *J Clin Virol* **37**(1) (2006), 21–26.
- [13] H.A. Cubie, M. Plumstead, W. Zhang, O. de Jesus, L.A. Duncan and M.A. Stanley, Presence of antibodies to human papillomavirus virus-like particles (VLPs) in 11–13-year-old schoolgirls, *J Med Virol* **56**(3) (1998), 210–216.
- [14] D.M. Da Silva, G.L. Eiben, S.C. Fausch, M.T. Wakabayashi, M.P. Rudolf, M.P. Velders and W.M. Kast, Cervical cancer vaccines: emerging concepts and developments, *J Cell Physiol* **186**(2) (2001), 169–182.
- [15] M. Dürst, L. Gissmann, H. Ikenberg and H. zur Hausen, A papillomavirus DNA from a cervical carcinoma and its prevalence in cancer biopsy samples from different geographic regions, *Proc Natl Acad Sci USA* **80**(12) (1983), 3812–3815.
- [16] G.L. Eiben, D.M. da Silva, S.C. Fausch, I.C. Le Poole, M.I. Nishimura and W.M. Kast, Cervical cancer vaccines: recent advances in HPV research, *Viral Immunol* **16**(2) (2003), 111–121.
- [17] R.T. Emeny, C.M. Wheeler, K.U. Jansen, W.C. Hunt, T.M. Fu, J.F. Smith, S. MacMullen, M.T. Esser and X. Paliard, Priming of human papillomavirus type 11-specific humoral and cellular immune responses in college-aged women with a virus-like particle vaccine, *J Virol* **76**(15) (2002), 7832–7842.
- [18] C.A. Evans, L.R. Gorman, Y. Ito and R.S. Weiser, A vaccination procedure which increases the frequency of regressions of Shope papillomas of rabbits, *Nature* **193** (1962), 288–289.
- [19] T.G. Evans, W. Bonnez, R.C. Rose, S. Koenig, L. Demeter, J.A. Suzich, D. O'Brien, M. Campbell, W.I. White, J. Balesley and R.C. Reichman, A Phase 1 study of a recombinant viruslike particle vaccine against human papillomavirus type 11 in healthy adult volunteers, *J Infect Dis* **183**(10) (2001), 1485–1493.
- [20] K.H. Fife, C.M. Wheeler, L.A. Koutsky, E. Barr, D.R. Brown, M.A. Schiff, N.B. Kiviat, K.U. Jansen, H. Barber, J.F. Smith, A. Tadesse, K. Giacoletti, P.R. Smith, G. Suhr and D.A. Johnson, Dose-ranging studies of the safety and immunogenicity of human papillomavirus Type 11 and Type 16 virus-like particle candidate vaccines in young healthy women, *Vaccine* **22**(21–22) (2004), 2943–2952.
- [21] G. Findlay, Warts, in: *A system of bacteriology in relation to medicine*, (Vol. 7), pp. 252–258. H. M. Stationary Office, London, 1930.
- [22] F. Gagnon, Contribution to the study of the etiology and prevention of cancer of the cervix of the uterus, *Am J Obstet Gynecol* **60**(3) (1950), 516–522.
- [23] L. Gissmann, V. Diehl, H.J. Schultz-Coulon and H. zur Hausen, Molecular cloning and characterization of human papilloma virus DNA derived from a laryngeal papilloma, *J Virol* **44**(1) (1982), 393–400.
- [24] L. Gissmann and H. zur Hausen, Partial characterization of viral DNA from human genital warts (*Condylomata acuminata*), *Int J Cancer* **25**(5) (1980), 605–609.
- [25] S.J. Goldie, M. Kohli, D. Grima, M.C. Weinstein, T.C. Wright, F.X. Bosch and E. Franco, Projected clinical benefits and cost-

- effectiveness of a human papillomavirus 16/18 vaccine, *J Natl Cancer Inst* **96**(8) (2004), 604–615.
- [26] W. Grigg and G. Wilhelm, Epidemiological study of plantar warts among school children, *Public Health Report US* **68** (1953), 985–988.
- [27] M.E. Hagensee, N.H. Olson, T.S. Baker and D.A. Galloway, Three-dimensional structure of vaccinia virus-produced human papillomavirus type 1 capsids, *J Virol* **68**(7) (1994), 4503–4505.
- [28] R. Han, N.M. Cladel, C.A. Reed, X. Peng and N.D. Christensen, Protection of rabbits from viral challenge by gene gun-based intracutaneous vaccination with a combination of cottontail rabbit papillomavirus E1, E2, E6, and E7 genes, *J Virol* **73**(8) (1999), 7039–7043.
- [29] R. Han, C.A. Reed, N.M. Cladel and N.D. Christensen, Immunization of rabbits with cottontail rabbit papillomavirus E1 and E2 genes: protective immunity induced by gene gun-mediated intracutaneous delivery but not by intramuscular injection, *Vaccine* **18**(26) (2000), 2937–2944.
- [30] D.M. Harper, E.L. Franco, C. Wheeler, D.G. Ferris, D. Jenkins, A. Schuind, T. Zahaf, B. Innis, P. Naud and N.S. De Carvalho, Efficacy of a bivalent L1 virus-like particle vaccine in prevention of infection with human papillomavirus types 16 and 18 in young women: a randomised controlled trial, *The Lancet* **364**(9447) (2004), 1757–1765.
- [31] C.D. Harro, Y.Y. Pang, R.B. Roden, A. Hildesheim, Z. Wang, M.J. Reynolds, T.C. Mast, R. Robinson, B.R. Murphy, R.A. Karron, J. Dillner, J.T. Schiller and D.R. Lowy, Safety and immunogenicity trial in adult volunteers of a human papillomavirus 16 L1 virus-like particle vaccine, *J Natl Cancer Inst* **93**(4) (2001), 284–292.
- [32] S. Inglis, A. Shaw and S. Koenig, Chapter 11: HPV vaccines: Commercial Research & Development, *Vaccine* **24**(Suppl 3) (2006), S99–S105.
- [33] Y. Ito and C.A. Evans, Tumorigenic Nucleic Acid Extracts from Tissues of a Transplantable Carcinoma, Vx7, *J Natl Cancer Inst* **34** (1965), 431–437.
- [34] W.F. Jarrett, B.W. O'Neil, J.M. Gaukroger, K.T. Smith, H.M. Laird and M.S. Campo, Studies on vaccination against papillomaviruses: the immunity after infection and vaccination with bovine papillomaviruses of different types, *Vet Rec* **126**(19) (1990), 473–475.
- [35] R. Kirnbauer, L.M. Chandrachud, B.W. O'Neil, E.R. Wagner, G.J. Grindlay, A. Armstrong, G.M. McGarvie, J.T. Schiller, D.R. Lowy and M.S. Campo, Virus-like particles of bovine papillomavirus type 4 in prophylactic and therapeutic immunization, *Virology* **219**(1) (1996), 37–44.
- [36] R. Kirnbauer, J. Taub, H. Greenstone, R. Roden, M. Dürst, L. Gissmann, D.R. Lowy and J.T. Schiller, Efficient self-assembly of human papillomavirus type 16 L1 and L1-L2 into virus-like particles, *J Virol* **67**(12) (1993), 6929–6936.
- [37] L.A. Koutsky, K.A. Ault, C.M. Wheeler, D.R. Brown, E. Barr, F.B. Alvarez, L.M. Chiacchierini and K.U. Jansen, A controlled trial of a human papillomavirus type 16 vaccine, *N Engl J Med* **347**(21) (2002), 1645–1651.
- [38] J.W. Kreider, M.K. Howett, A.E. Leure-Dupree, R.J. Zaino and J.A. Weber, Laboratory production *in vivo* of infectious human papillomavirus type 11, *J Virol* **61**(2) (1987), 590–593.
- [39] E. Lazcano-Ponce, L. Rivera, E. Arillo-Santillan, J. Salmeron, M. Hernandez-Avila and N. Munoz, Acceptability of a human papillomavirus (HPV) trial vaccine among mothers of adolescents in Cuernavaca, Mexico, *Arch Med Res* **32**(3) (2001), 243–247.
- [40] M. Lehtinen and J. Paavonen, Effectiveness of preventive human papillomavirus vaccination, *Int J STD AIDS* **14**(12) (2003), 787–792.
- [41] W. Licht, Om Vorters Smitsomhed, *Ugeskrift Laeger* **1** (1894), 368–369.
- [42] R.S. Lowe, D.R. Brown, J.T. Bryan, J.C. Cook, H.A. George, K.J. Hofmann, W.M. Hurni, J.G. Joyce, E.D. Lehman, H.Z. Markus, M.P. Neepser, L.D. Schultz, A.R. Shaw and K.U. Jansen, Human papillomavirus type 11 (HPV-11) neutralizing antibodies in the serum and genital mucosal secretions of African green monkeys immunized with HPV-11 virus-like particles expressed in yeast, *J Infect Dis* **176**(5) (1997), 1141–1145.
- [43] D. Lowy and P. Howley, Papillomaviruses, in: *Fields Virology*, (Vol. 1), D. Knipe and P. Howley, eds, 2001, pp. 2231–2264. 2 vols. Lippincott, Williams and Wilkins, Philadelphia, USA.
- [44] C. Mao, L.A. Koutsky, K.A. Ault, C.M. Wheeler, D.R. Brown, D.J. Wiley, F.B. Alvarez, O.M. Bautista, K.U. Jansen and E. Barr, Efficacy of Human Papillomavirus-16 Vaccine to Prevent Cervical Intraepithelial Neoplasia: A Randomized Controlled Trial, *Obstet Gynecol* **107**(1) (2006), 18–27.
- [45] A.M. Massing and W.L. Epstein, Natural history of warts. A two-year study, *Arch Dermatol* **87** (1963), 306–310.
- [46] R.M. Mays, L.A. Sturm and G.D. Zimet, Parental perspectives on vaccinating children against sexually transmitted infections, *Soc Sci Med* **58**(7) (2004), 1405–1413.
- [47] T.J. Palker, J.M. Monteiro, M.M. Martin, C. Kakareka, J.F. Smith, J.C. Cook, J.G. Joyce and K.U. Jansen, Antibody, cytokine and cytotoxic T lymphocyte responses in chimpanzees immunized with human papillomavirus virus-like particles, *Vaccine* **19**(27) (2001), 3733–3743.
- [48] D.V. Pastrana, C.B. Buck, Y.Y. Pang, C.D. Thompson, P.E. Castle, P.C. FitzGerald, S. Kruger Kjaer, D.R. Lowy and J.T. Schiller, Reactivity of human sera in a sensitive, high-throughput pseudovirus-based papillomavirus neutralization assay for HPV16 and HPV18, *Virology* **321**(2) (2004), 205–216.
- [49] I. Penn, Cancers of the anogenital region in renal transplant recipients. Analysis of 65 cases, *Cancer* **58**(3) (1986), 611–616.
- [50] W.P. Pilacinski, D.L. Glassman, K.F. Glassman, D.E. Reed, M.A. Lum, R.F. Marshall, C.C. Muscoplat and A.J. Faras, Immunization against bovine papillomavirus infection, *Ciba Found Symp* **120** (1986), 136–156.
- [51] Rigoni-Stern, Fatti statistici relativi alle malattie cancerose, *G Serve Prog Pathol Terap* **2** (1842), 507–517.
- [52] R.B. Roden, N.L. Hubbert, R. Kirnbauer, F. Breitburd, D.R. Lowy and J.T. Schiller, Papillomavirus L1 capsids agglutinate mouse erythrocytes through a proteinaceous receptor, *J Virol* **69**(8) (1995), 5147–5151.
- [53] R.B. Roden, E.M. Weissinger, D.W. Henderson, F. Booy, R. Kirnbauer, J.F. Mushinski, D.R. Lowy and J.T. Schiller, Neutralization of bovine papillomavirus by antibodies to L1 and L2 capsid proteins, *J Virol* **68**(11) (1994), 7570–7574.
- [54] R.C. Rose, R.C. Reichman and W. Bonnez, Human papillomavirus (HPV) type 11 recombinant virus-like particles induce the formation of neutralizing antibodies and detect HPV-specific antibodies in human sera, *J Gen Virol* **75**(Pt 8) (1994), 2075–2079.
- [55] M.P. Rudolf, S.C. Fausch, D.M. Da Silva and W.M. Kast, Human dendritic cells are activated by chimeric human papillomavirus type-16 virus-like particles and induce epitope-specific human T cell responses *in vitro*, *J Immunol* **166**(10) (2001), 5917–5924.

- [56] D.M. Salunke, D.L. Caspar and R.L. Garcea, Self-assembly of purified polyomavirus capsid protein VP1, *Cell* **46**(6) (1986), 895–904.
- [57] K. Schafer, M. Müller, S. Faath, A. Henn, W. Osen, H. Zentgraf, A. Benner, L. Gissmann and I. Jochmus, Immune response to human papillomavirus 16 L1E7 chimeric virus-like particles: induction of cytotoxic T cells and specific tumor protection, *Int J Cancer* **81**(6) (1999), 881–888.
- [58] J.T. Schiller and D.R. Lowy, Papillomavirus-like particle vaccines, *J Natl Cancer Inst Monogr* (28) (2001), 50–54.
- [59] E. Schwarz, M. Dürst, C. Demankowski, O. Lattermann, R. Zech, E. Wolfsperger, S. Suhai and H. zur Hausen, DNA sequence and genome organization of genital human papillomavirus type 6b, *Embo J* **2**(12) (1983), 2341–2348.
- [60] R. Shope, Infectious papillomatosis of rabbits; with a note on histopathology, *J Exp Med* **58** (1933), 607–624.
- [61] M.A. Stanley, Progress in prophylactic and therapeutic vaccines for human papillomavirus infection, *Expert Rev Vaccines* **2**(3) (2003), 381–389.
- [62] R.S. Taylor, B.E. Carroll and J.W. Lloyd, Mortality among women in 3 Catholic religious orders with special reference to cancer, *Cancer* **12** (1959), 1207–1225.
- [63] J.E. Towne, Carcinoma of the cervix in nulliparous and celibate women, *Am J Obstet Gynecol* **69**(3) (1955), 606–613.
- [64] G. Variot, Un cas d'inoculation experimentale des verruces de l'enfant a l'homme, *J Clin Therapy Infant* **2** (1894), 529–534.
- [65] L.L. Villa, R.L. Costa, C.A. Petta, R.P. Andrade, K.A. Ault, A.R. Giuliano, C.M. Wheeler, L.A. Koutsky, C. Malm, M. Lehtinen, F.E. Skjeldestad, S.E. Olsson, M. Steinwall, D.R. Brown, R.J. Kurman, B.M. Ronnett, M.H. Stoler, A. Ferenczy, D.M. Harper, G.M. Tamms, J. Yu, L. Lupinacci, R. Railkar, F.J. Taddeo, K.U. Jansen, M.T. Esser, H.L. Sings, A.J. Saah and E. Barr, Prophylactic quadrivalent human papillomavirus (types 6, 11, 16, and 18) L1 virus-like particle vaccine in young women: a randomised double-blind placebo-controlled multicentre phase II efficacy trial, *Lancet Oncol* **6**(5) (2005), 271–278.
- [66] V. Vonka, J. Kanka, J. Jelinek, I. Subrt, A. Suchanek, A. Havrankova, M. Vachal, I. Hirsch, E. Domorazkova, H. Zavadova et al., Prospective study on the relationship between cervical neoplasia and herpes simplex type-2 virus. I. Epidemiological characteristics, *Int J Cancer* **33**(1) (1984), 49–60.
- [67] L. Waelsch, Übertragungsversuche mit spitzen Kondyloma, *Arch Dermatol Syphilis* **124** (1918), 625–646.
- [68] J.M. Walboomers, M.V. Jacobs, M.M. Manos, F.X. Bosch, J.A. Kummer, K.V. Shah, P.J. Snijders, J. Peto, C.J. Meijer and N. Munoz, Human papillomavirus is a necessary cause of invasive cervical cancer worldwide, *J Pathol* **189**(1) (1999), 12–19.
- [69] J. Zhou, D.J. Stenzel, X.Y. Sun and I.H. Frazer, Synthesis and assembly of infectious bovine papillomavirus particles *in vitro*, *J Gen Virol* **74** (Pt 4) (1993), 763–768.
- [70] J. Zhou, X.Y. Sun, H. Davies, L. Crawford, D. Park and I.H. Frazer, Definition of linear antigenic regions of the HPV16 L1 capsid protein using synthetic virion-like particles, *Virology* **189**(2) (1992), 592–599.
- [71] G.D. Zimet, R.M. Mays, Y. Winston, R. Kee, J. Dickes and L. Su, Acceptability of human papillomavirus immunization, *J Womens Health Gend Based Med* **9**(1) (2000), 47–50.
- [72] H. zur Hausen, Condylomata acuminata and human genital cancer, *Cancer Res* **36**(2 pt 2) (1976), 794.



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