

## Research Article

# Thyroxine Alleviates Energy Failure, Prevents Myocardial Cell Apoptosis, and Protects against Doxorubicin-Induced Cardiac Injury and Cardiac Dysfunction via the LKB1/AMPK/mTOR Axis in Mice

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**Background.** Previous studies have demonstrated that energy failure is closely associated with cardiac injury. Doxorubicin (DOX) is a commonly used clinical chemotherapy drug that can mediate cardiac injury through a variety of mechanisms. Thyroxine is well known to play a critical role in energy generation; thus, this study is aimed at investigating whether thyroxine can attenuate DOX-induced cardiac injury by regulating energy generation. **Methods.** First, the effect of DOX on adenosine diphosphate (ADP) and adenosine triphosphate (ATP) ratios in mice was assessed. In addition, thyroxine was given to mice before they were treated with DOX to investigate the effects of thyroxine on DOX-induced cardiac injury. Furthermore, to determine whether the liver kinase b1 (LKB1)/adenosine 5'-monophosphate-activated protein kinase (AMPK)/mammalian target of rapamycin (mTOR) axis mediated the effect of thyroxine on DOX-induced cardiac injury, thyroxine was given to DOX-treated LKB1 knockout (KO) mice. **Results.** DOX treatment time- and dose-dependently increased the ADP/ATP ratio. Thyroxine treatment also reduced lactate dehydrogenase (LDH) and creatine kinase myocardial band (CK-MB) levels in both serum and heart tissue and alleviated cardiac dysfunction in DOX-treated mice. Furthermore, thyroxine reversed DOX-induced reductions in LKB1 and AMPK phosphorylation; mitochondrial complex I, III, and IV activity; and mitochondrial swelling and reversed DOX-induced increases in mTOR pathway phosphorylation and myocardial cell apoptosis. These effects of thyroxine on DOX-treated mice were significantly attenuated by LKB1 KO. **Conclusions.** Thyroxine alleviates energy failure, reduces myocardial cell apoptosis, and protects against DOX-induced cardiac injury via the LKB1/AMPK/mTOR axis in mice. Thyroxine may be a new agent for the clinical prevention of cardiac injury in tumor patients undergoing chemotherapy with DOX.

## 1. Introduction

Doxorubicin (DOX) is a periodic nonspecific antitumor drug that can kill different tumor cells in various growth stages and is widely used in clinical chemotherapy for numerous types of clinical tumors [1]. Although DOX has excellent antitumor effects, it inevitably leads to a variety of serious clinical

complications, especially cardiac injury, which may pose a more serious threat to the patient's life than the tumor does [2, 3]. Because of the large population of tumor patients and the widespread use of DOX among them, finding ways to combat DOX-induced cardiac injury is of great social value.

The human body is a complex metabolic factory, and a normal energy supply is essential to the proper functioning

of all organs, especially the heart and brain [4]. Growing evidence has demonstrated that decreases in energy production from various causes are closely related to the occurrence of cardiac injury, including PIMT/NCOA6IP gene deletion-induced delayed cardiomyopathy, diabetic cardiomyopathy, and global ischemia-induced cardiac injury [5–7]. The underlying mechanisms may be related to excessive apoptosis caused by the energy generation of cardiac cells [8]. DOX-induced cardiac injury is a complex pathophysiological process in which a variety of mechanisms have been demonstrated to be involved, including excessive cardiac inflammatory responses, oxidative stress, autophagy, and apoptosis [9]. However, whether abnormal energy metabolism is involved in the progression of DOX-induced cardiac injury remains unclear.

Thyroxine, an essential hormone, has been linked to growth, apoptosis, and energy metabolism and has been demonstrated to be closely associated with cardiovascular diseases. It is well known that prolonged elevations in thyroxine levels in patients with hyperthyroidism can lead to changes in cardiac structure. A recent study reported that among female South Korean patients over 30 years of age, subclinical hypothyroidism significantly increased the 10-year cardiovascular disease risk score [10]. Almost at the same time, another article reported that thyroxine could regulate lipid status and affect the occurrence of cardiovascular events, although this phenomenon was not observed in patients with subclinical hypothyroidism [11]. In another clinical experiment, the authors found that increased thyroxine levels significantly increased the incidence of atherosclerotic cardiovascular (ASCV) events and ASCV mortality [12]. However, whether thyroxine is involved in energy metabolism and cardiac injury remains unknown. We hypothesized that thyroxine could alleviate DOX-induced cardiac injury via the regulation of abnormal energy metabolism. In the present study, we performed the following experiments to confirm this speculation.

## 2. Methods and Materials

**2.1. Animals and Animal Models.** Both male wild-type (WT) mice and liver kinase b1- (LKB1-) knockout (KO) mice on a C57BL/6 background (aged 10–11 weeks) were used in this study (both purchased from the Model Animal Research Center of Nanjing University, China). First, the WT mice received an intraperitoneal (i.p.) injection of different doses (5 mg/kg, 10 mg/kg, or 15 mg/kg) of DOX (purity  $\geq$  98%) for 5 days or were treated with 15 mg/kg DOX for different lengths of time (1 day, 3 days, or 5 days); WT mice received the same amount of saline as the control (baseline group). The adenosine diphosphate/adenosine triphosphate (ADP/ATP) ratio in each mouse was assessed ( $n = 6$  for each group). In addition, WT mice also received phosphate-buffered saline (PBS) (50  $\mu$ l) or thyroxine (1.25 mg/kg) treatment daily for 3 days [13] and then were given DOX; WT mice that received PBS were treated with saline as controls ( $n = 12$  for each group). The body weight of each mouse was recorded daily before and after DOX treatment (15 mg/kg). Five days later, the mice were eutha-

nized after echocardiography, hemodynamic analyses were performed, and blood samples and heart samples were collected for subsequent analysis. In a separate experiment, both WT mice and LKB1-KO mice were treated daily with PBS or thyroxine. Three days later, all the mice were given DOX ( $n = 12$  for each group), and heart tissue was also collected. All mice were housed in the specific-pathogen-free mouse room of Renmin Hospital of Wuhan University, and this study was reviewed and approved by the Institutional Animal Care and Use Committee at Renmin Hospital, Wuhan University (China).

**2.2. Echocardiography and Hemodynamics.** Isoflurane at a 2% concentration delivered through inhalation was used to anesthetize the mice. Both a MyLab 30CV ultrasound (Esaote SpA, Italy) system and a 10 MHz linear array ultrasound transducer were used to perform echocardiography. M-mode images of the left ventricle (LV) at the papillary muscle level were recorded, and parameters related to LV structure and function, including heart rate (HR), left ventricular ejection fraction (LVEF), and fractional shortening (FS), were measured. For the hemodynamic analysis, a microtip catheter transducer (Millar, USA) was inserted into the LV through the right carotid artery, and a Millar Pressure-Volume System (Millar) was used to record continuous signals, such as the maximal slope of the systolic pressure increase (+dp/dt max) and diastolic pressure decrease (-dp/dt max), left ventricular systolic pressure (LVSP), and left ventricular end-diastolic pressure (LVEDP), on a beat-by-beat basis.

**2.3. Measurement of ADP/ATP Ratio.** LV tissue was lysed with self-contained lysate, and the supernatant of each sample was collected. Both the ADP and the ATP levels in each sample were measured using an ADP assay kit and an ATP assay kit, respectively, following the manufacturer's instructions (both kits were from Beyotime, China).

**2.4. Histological Analysis.** After isolation and weighing, the heart was immediately fixed with 4% neutral paraformaldehyde for 5 days, embedded in paraffin, and cut into approximately 4–7  $\mu$ m sections. The sections were mounted onto slides, and hematoxylin and eosin (HE) staining was performed. After HE staining was completed and the sections were photographed, the number of cardiomyocytes and the number of vacuolated cardiomyocytes, which is a pathological change representing degeneration and necrosis occurring in the cardiomyocytes, are shown. The ratio of vacuolated cardiomyocytes to total cardiomyocytes was calculated for each heart sample. Apoptosis was detected using a terminal deoxynucleotidyl transferase-mediated dUTP nick end-labeling (TUNEL) staining kit (Millipore, USA) according to the manufacturer's instructions. In addition, the morphology of the mitochondria was examined by transmission electron microscopy.

**2.5. Measurement of Lactate Dehydrogenase (LDH) and Creatine Kinase Myocardial Band (CK-MB).** After blood samples were centrifuged for 15 min at 4000  $\times$ g, serum was obtained, and supernatant was collected from LV tissue.

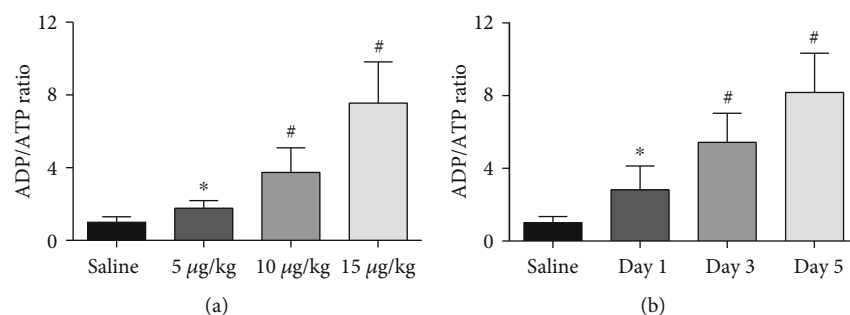


FIGURE 1: Cardiac mitochondrial ADP/ATP ratios in saline- and DOX-treated mice. (a) The effects of different DOX doses on cardiac mitochondrial ADP/ATP ratios were determined. (b) The effects of DOX on cardiac mitochondrial ADP/ATP ratios at different time points were measured.  $N = 6$  in each group. \* $P < 0.05$  vs. saline group; # $P < 0.01$  vs. previous concentration and time point.

LDH and CK-MB levels in both serum and supernatant were determined with kits according to the manufacturer's instructions (both kits from Beyotime, China).

**2.6. Western Blot Analysis.** Radioimmunoprecipitation assay (RIPA) lysate buffer containing protease inhibitors and phosphatase inhibitors was used to lyse LV tissue, and the total protein in each sample was collected and quantified with a BCA Protein Assay Kit (Thermo Fisher Scientific, USA). Electrophoresis was performed to separate the total protein using 10% sodium dodecyl sulfate (SDS) polyacrylamide gels, and the separated proteins were then transferred to polyvinylidene difluoride (PVDF) membranes (Millipore, USA). After blocking with 5% nonfat milk for 1 h, the membranes were incubated with primary antibodies at 4°C overnight and then incubated with secondary antibodies at room temperature for 1 h. The primary antibodies included anti-LKB1, anti-p-LKB1, anti-adenosine 5'-monophosphate-activated protein kinase (AMPK), anti-p-AMPK, anti-mammalian target of rapamycin (mTOR), anti-p-mTOR, anti-Bax, anti-Bcl2, anti-cleaved caspase-3, and anti-GAPDH antibodies (all from Cell Signaling Technology). The blots were scanned using a two-color infrared imaging system (Odyssey, USA).

**2.7. Mitochondrial Complex I, III, and IV Activity Assays.** Mitochondria were isolated from fresh LV samples as previously described [14]. Complex I, III, and IV assay kits were used to detect the specific activity of mitochondrial complexes I, III, and IV, respectively, according to the manufacturer's instructions (all three kits were from GENMED Scientifics, China). Mitochondrial complex I is nicotinamide adenine dinucleotide Q reductase, mitochondrial complex III is coenzyme Q-cytochrome c reductase, and mitochondrial complex IV is cytochrome c oxidase. The activity of each mitochondrial complex is expressed as nanomoles per minute per milligram of protein.

**2.8. Statistical Analyses.** All data in this study are expressed as the mean  $\pm$  standard deviation (SD). Student's  $t$ -test and one-way analysis of variance (ANOVA) followed by Tukey's multiple comparison test were used to compare differences between the means of two groups and among the means of

more than two groups, respectively. A value of  $P < 0.05$  was considered significant.

### 3. Results

**3.1. DOX Treatment Increases the ADP/ATP Ratio.** The mitochondrial ADP/ATP ratio can be used to reflect energy failure. To clarify the effect of DOX on cardiac energy failure, the mitochondrial ADP/ATP ratio was first detected. The results showed that compared with the baseline levels, the ADP/ATP ratios were dose-dependently increased by DOX treatment (Figure 1(a)). Similar results were observed when the mice were treated with DOX for different durations (Figure 1(b)).

**3.2. Thyroxine Treatment Alleviates DOX-Induced Cardiac Injury.** To determine the effect of thyroxine on DOX-induced cardiac injury, we first examined its effect on body weight and heart weight. The results showed that DOX treatment for 5 days reduced body weight by approximately 4.8 g; however, this DOX-induced reduction in body weight was reversed by thyroxine treatment, as indicated by the fact that only 2.9 g of body weight loss was observed in the DOX+thyroxine group (Figure 2(a)). Thyroxine treatment also significantly alleviated heart weight loss in DOX-treated mice (Figure 2(b)). Vacuolation is also an important manifestation of myocardial injury, and the trends in the numbers of vacuolated cardiomyocytes were similar to those observed for body weight and heart weight (Figure 2(c)). In addition, LDH and CK-MB can indicate the degree of heart damage; therefore, thyroxine was also observed to decrease the LDH and CK-MB levels in both serum and heart tissue from DOX-treated mice (Figures 2(d)–2(g)).

**3.3. Thyroxine Treatment Relieves DOX-Induced Cardiac Dysfunction.** The results of echocardiography revealed lower HR, LVEF, and FS in the DOX group than in the control group, but these decreases were partly prevented by thyroxine administration in DOX-treated mice (Figures 3(a)–3(c)). Similar results were obtained from invasive hemodynamic measurements. Both systolic (Figures 3(d) and 3(f)) and diastolic (Figures 3(e) and 3(g)) LV dysfunctions induced by DOX were reduced by thyroxine treatment.

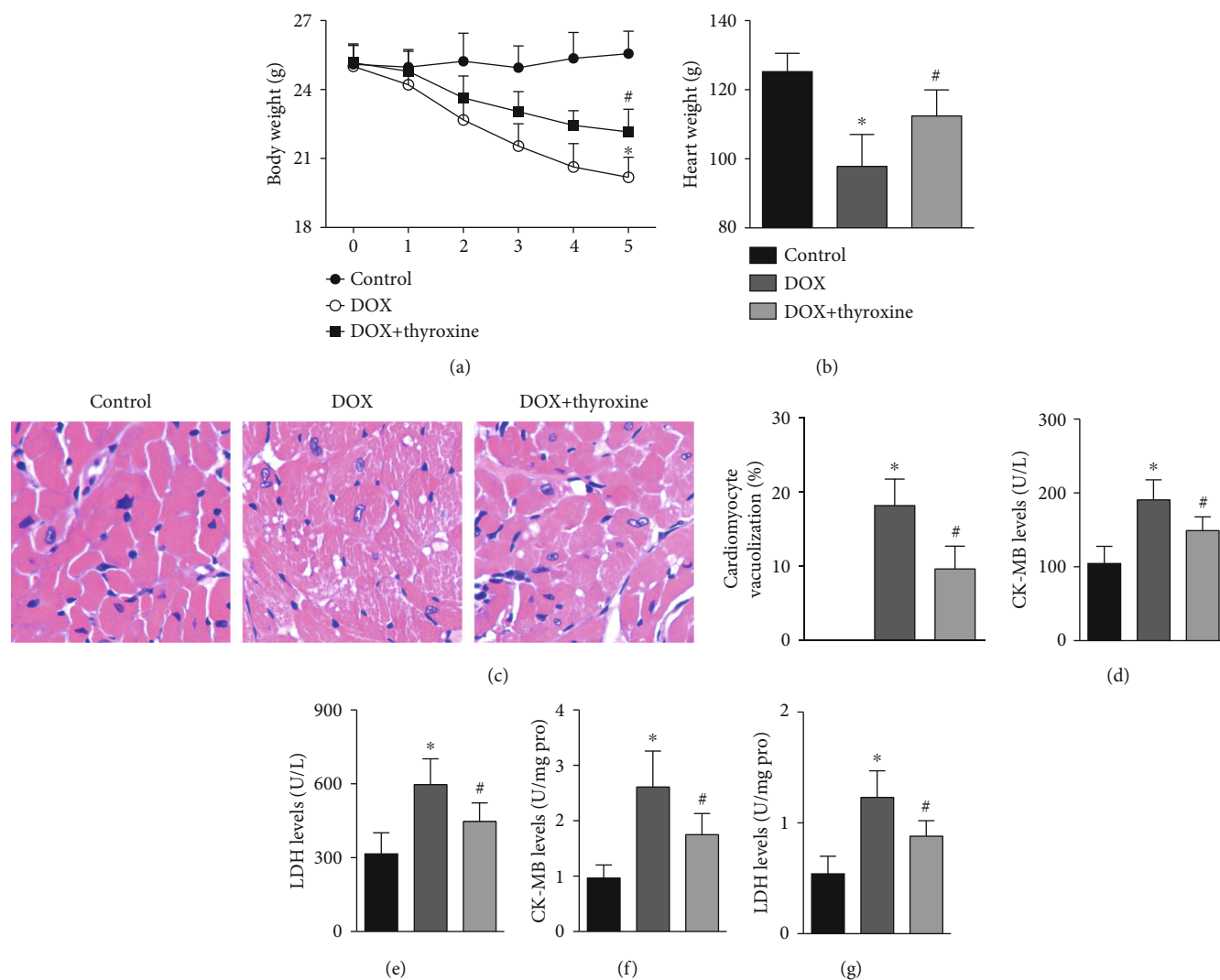


FIGURE 2: Effects of thyroxine on DOX-induced cardiac injury. (a) Body weights in the four groups were measured at different time points;  $N = 12$  in each group. (b) Heart weights were measured at the end of the fifth day in each group;  $N = 12$  in each group. (c) Heart sections were stained with HE, and the numbers of vacuolated cardiomyocytes were quantified (200 $\times$ );  $N = 6$  in each group. (d, e) Serum LDH and CK-MB levels were assessed;  $N = 12$  in each group. (f, g) Heart LDH and CK-MB levels were detected;  $N = 6$  in each group. \* $P < 0.05$  vs. control group; # $P < 0.01$  vs. DOX group.

**3.4. Thyroxine Treatment Decreases DOX-Induced Reductions in Energy Generation.** The LKB1/AMPK/mTOR axis is closely related to energy production. To investigate the effects of thyroxine on LKB1/AMPK/mTOR axis activation, the LKB1, AMPK, and mTOR phosphorylation in LV was measured, and the results showed that DOX treatment significantly decreased LKB1 and AMPK phosphorylation while increasing mTOR phosphorylation. Both the decreased LKB1 and AMPK phosphorylation and the increased mTOR phosphorylation were partially reversed by thyroxine in DOX-treated mice (Figure 4(a)). DOX treatment also reduced mitochondrial complex I, III, and IV activation, and these effects were reversed by thyroxine treatment (Figure 4(b)). In addition, DOX-induced mitochondrial swelling was also prevented when thyroxine was given (Figure 4(c)).

**3.5. Thyroxine Treatment Reduces DOX-Induced Myocardial Apoptosis.** The essence of DOX-induced cardiac injury is

excessive apoptosis of cardiomyocytes; therefore, the effect of thyroxine on cardiomyocyte apoptosis was examined. The results showed that 5 days after DOX treatment, the levels of Bcl2 were lower, and the levels of Bax and cleaved caspase-3 were higher in the DOX group than in the control group (Figure 5(a)). Thyroxine treatment decreased Bax and cleaved caspase-3 levels and increased Bcl2 levels in DOX-treated mice (Figure 5(b)). In addition, thyroxine significantly reduced the DOX-induced increases in the number of TUNEL-positive cells in DOX-treated mice (Figure 5(b)).

**3.6. LKB1 KO Alleviates the Effects of Thyroxine on DOX-Induced Cardiac Injury.** To test whether thyroxine regulates the effects of DOX-induced cardiac injury via the LKB1/AMPK/mTOR axis, LKB1 was knocked out, and the effects on downstream signals were detected. The thyroxine-mediated reduction in DOX-induced vacuolated cardiomyocytes was significantly reversed by LKB1 KO



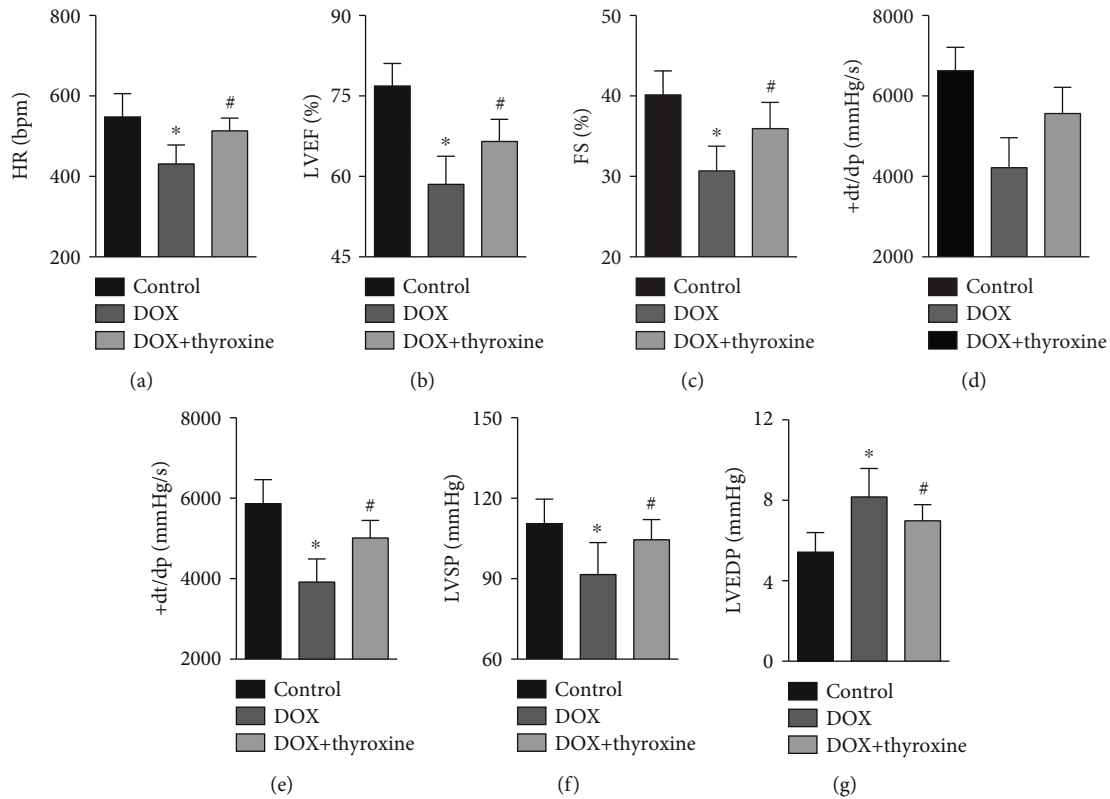


FIGURE 3: Left ventricular function was measured by echocardiography and analysis of hemodynamic data in mice treated with PBS or thyroxine (1.25 mg/kg) after administration of saline or DOX (15 mg/kg) for 5 days. (a–c) HR, LVEF, and FS were evaluated in the three groups by echocardiography. (d–g) Positive and negative dP/dt max, LVSP, and LVEDP were assessed in each group based on hemodynamic data.  $N = 12$  in each group. \* $P < 0.05$  vs. control group; # $P < 0.01$  vs. DOX group.

(Figure 6(a)). LKB1 KO also increased heart LDH and CK-MB levels in DOX+thyroxine mice (Figure 6(b)). Furthermore, LKB1 KO decreased AMPK phosphorylation while increasing mTOR phosphorylation (Figure 6(c)). Regarding the expression of apoptosis-related proteins, higher Bax and cleaved caspase-3 levels and lower Bcl2 levels were observed in DOX+thyroxine mice when LKB1 was knocked out than when LKB1 was functional (Figure 6(d)). In addition, LKB1 KO prevented thyroxine-induced increases in mitochondrial complex I, III, and IV activation in DOX-treated mice (Figure 6(e)). The protective effect of thyroxine on DOX-induced mitochondrial swelling was also reversed by LKB1 KO (Figure 6(f)). Finally, LKB1 KO increased the number of TUNEL-positive cells in DOX+thyroxine mice (Figure 6(g)).

#### 4. Discussion

In the present study, we observed that DOX treatment for 5 days significantly increased the ADP/ATP ratios in mice. In addition, thyroxine activated the LKB1 and AMPK pathways while inhibiting mTOR phosphorylation and increasing mitochondrial complex I, III, and IV activation; reversed energy failure and mitochondrial swelling; alleviated cardiac injury; and improved cardiac dysfunction. The protective role of thyroxine against cardiac injury and dysfunction was prevented by LKB1 KO.

DOX is a powerful chemotherapy drug that can also cause damage to myocardial cells through a complex set of physiological effects when tumor cells are killed. The currently known mechanisms involved in DOX-induced cardiac injury include serious inflammatory responses, oxidative stress imbalance, severe myocardial cell autophagy, and apoptosis [9, 15–18]. The literature overwhelmingly suggests that myocardial cells can be directly damaged by an excessive inflammatory response, which exacerbates oxidative stress [9, 15, 19–21], while severe or rapid autophagy can induce programmed myocardial cell death and decrease myocardial cell survival; however, autophagy has dual effects on myocardial cells, as mild autophagy partly protects cells from harmful conditions and promotes cell survival [22, 23]. In a recent study, Räsänen et al. reported that cardiac vascular endothelial dysfunction and associated myocardial ischemia are also involved in DOX-induced cardiac injury [24]. In the present study, we first assessed the ADP/ATP ratios in DOX-treated mice and found that DOX treatment increased the ADP/ATP ratio. Considering the important indicated role of increased ADP/ATP ratios in reactive cardiomyocyte energy failure, these data suggest that DOX treatment could lead to energy failure. Given our above description of the association of energy supply dysfunction with multiple heart diseases, we suspect that energy failure may participate in the progression and development of DOX-induced myocardial injury.

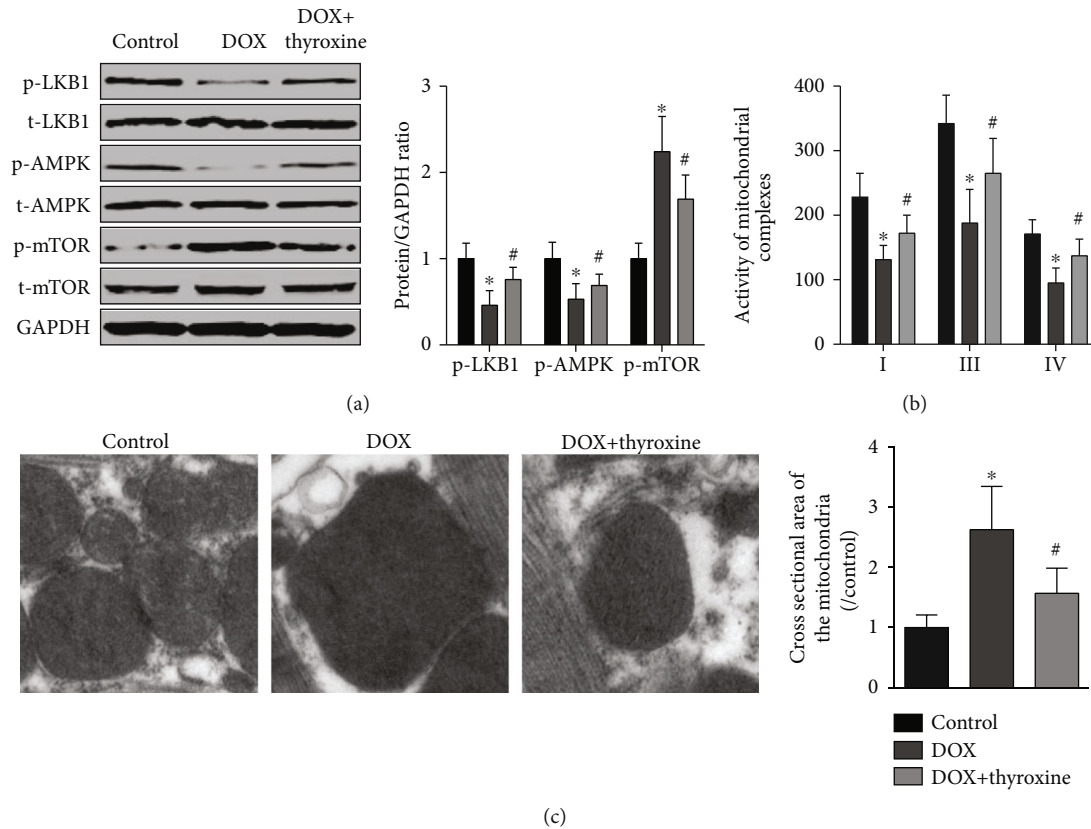


FIGURE 4: Effects of thyroxine on LKB1/AMPK/mTOR axis phosphorylation and mitochondrial complex I, III, and IV activation. (a) Representative images and quantification of p-LKB1, t-LKB1, p-AMPK, t-AMPK, p-mTOR, t-mTOR, and GAPDH in the LV for each group by western blot. (b) The activation of mitochondrial complexes I, III, and IV in the three groups was determined. (c) The cross-sectional area of the mitochondria in each group and more than 50 mitochondria were analyzed (5000x).  $N = 6$  in each group. \* $P < 0.05$  vs. control group; # $P < 0.01$  vs. DOX group.

To determine whether thyroxine could regulate DOX-induced cardiac injury, sufficient doses of thyroxine were given to mice prior to DOX treatment. In the studies that followed, we found that thyroxine significantly reduced the number of vacuolated cardiomyocytes and the levels of myocardial injury markers in DOX-treated mice. In addition, both the systolic and the diastolic dysfunctions induced by DOX were significantly reversed by thyroxine. These results demonstrated that thyroxine treatment alleviated DOX-induced cardiac injury and dysfunction.

Under the stimulation of various factors, regulation of cardiac metabolic demand is crucial to maintain normal cardiac function [4]. The AMPK pathway is considered to be the master regulator of cellular energy during conditions of metabolic stress because it can establish ATP homeostasis by both inhibiting ATP-consuming anabolic pathways and switching on catabolic pathways [25, 26]. LKB1 is the upstream signal of AMPK and is one of the most important kinases mediating AMPK phosphorylation [7]. While the mTOR pathway is also an upstream signal of AMPK, the LKB1/AMPK pathway can negatively regulate the phosphorylation of the mTOR pathway [7]. Considering the regulation of energy metabolism also reported in a recent study [7], we measured the effects of thyroxine on the phos-

phorylation of the LKB1/AMPK/mTOR axis to determine the mechanisms by which thyroxine alleviates DOX-induced cardiac injury, and the results showed that thyroxine increased LKB1/AMPK phosphorylation but reduced mTOR phosphorylation in DOX-treated mice. The LKB1/AMPK/mTOR axis has been reported to regulate multiple biological processes, such as apoptosis and tumor growth; therefore, we can conclude only that thyroxine may reduce DOX-induced cardiac injury by regulating energy metabolism.

ATP is one of the main sources of energy and is the most direct source of energy. Mitochondria are the most important sites of energy generation in eukaryotes, and most ATP is produced in the inner mitochondrial membrane. Two of the important oxidative respiratory chains are present in mitochondria, and thyroxine has been reported to regulate the generation of ATP via the first oxidative respiratory chain and to activate mitochondrial complexes I, III, and IV [27]. To investigate whether thyroxine relieves cardiac injury by regulating energy metabolism, the activation of mitochondrial complexes I, III, and IV was measured, and the results showed that DOX-induced inhibition of mitochondrial complex I, III, and IV activation was significantly reversed by thyroxine treatment. In addition, maintaining the normal function of mitochondria is also crucial for ATP production,

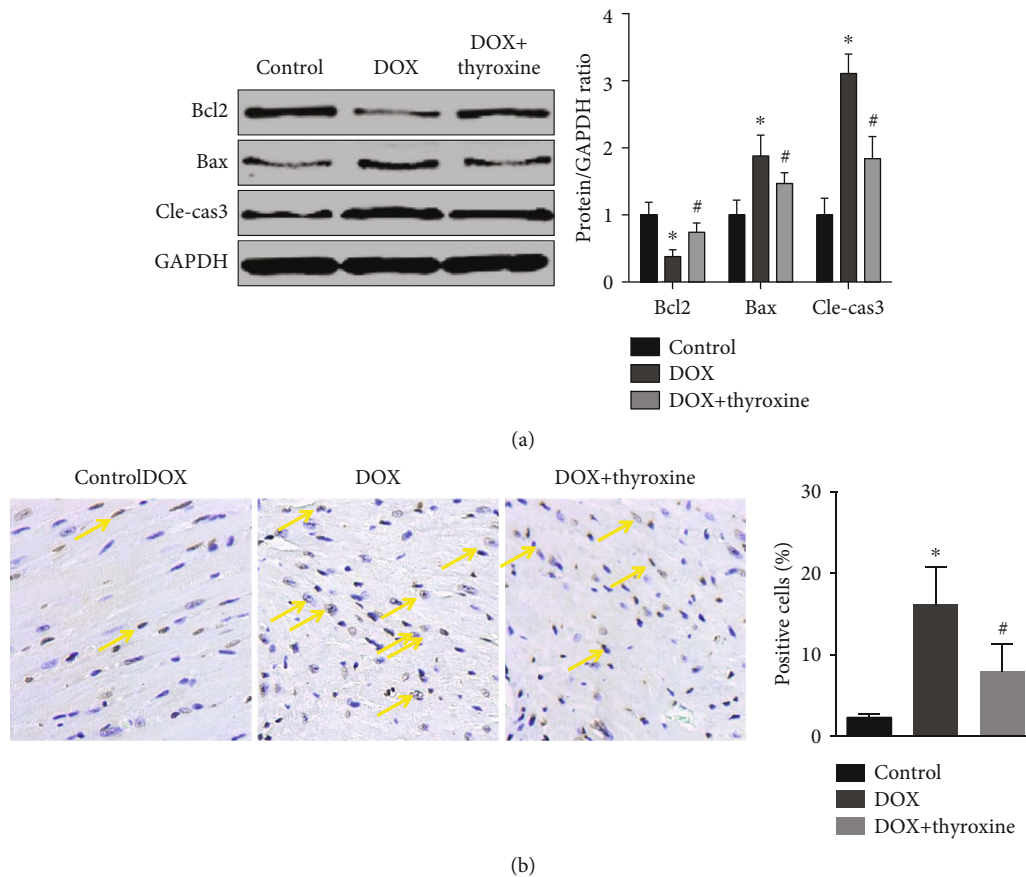


FIGURE 5: Effects of thyroxine on myocardial cell apoptosis. (a) Representative images and quantification of Bcl2, Bax, cleaved caspase-3 (cle-cas3), and GAPDH in the LV for each group by western blot. (b) Representative images of TUNEL staining and the quantitative results for each group (200x); arrows indicate TUNEL-positive cells.  $N = 6$  in each group. \* $P < 0.05$  vs. control group; # $P < 0.01$  vs. DOX group.

and its decline can lead to a significant decrease in ATP production. Mitochondrial dysfunction can cause structural changes, and swelling is one of the most important manifestations. Therefore, we examined the structure of the mitochondria and found that DOX led to mitochondrial swelling and that thyroxine reversed these effects. Our results are consistent with those of previous studies and may suggest that thyroxine protects against DOX-induced energy failure by stimulating the activation of mitochondrial complexes I, III, and IV and mitochondrial dysfunction.

The essence of cardiac injury induced by DOX is a change in cardiac structure and a decline in cardiac function caused by excessive apoptosis of myocardial cells [9]. In the present study, we also investigated the effect of thyroxine on myocardial cell apoptosis and found that Bax and cleaved caspase-3 protein levels and the number of TUNEL-positive cells were lower in the DOX+thyroxine group than in the DOX group. These results suggest that inhibition of LKB1/AMPK phosphorylation is involved in myocardial cell apoptosis. In fact, the effect of AMPK phosphorylation on myocardial cell apoptosis is controversial, and both a protective role and a pathogenic role were reported in a previous study [7, 28, 29]; the specific effects may be related to the length of the stimulus and the microenvironment of the organism.

As described above, mitigative effects on both energy failure and myocardial apoptosis were the mechanisms underlying thyroxine-mediated protection against DOX-induced cardiac injury. To determine whether these effects were mediated by the LKB1/AMPK/mTOR axis, LKB1 KO mice were given thyroxine and further treated with DOX. The results showed that LKB1 KO significantly decreased myocardial cell injury marker levels and prevented both the increased AMPK phosphorylation and decreased mTOR phosphorylation mediated by thyroxine in DOX-treated mice. In addition, the thyroxine-induced activation of mitochondrial complexes I, III, and IV and the protective effect of thyroxine in myocardial cells were also inhibited by LKB1 KO.

In summary, we found that energy failure was another mechanism that could participate in the progression and development of DOX-induced cardiac injury and has not been reported in previous studies. In addition, we found that thyroxine activated the LKB1/AMPK pathway and inhibited the mTOR pathway, alleviated energy failure, reduced myocardial cell apoptosis, and prevented DOX-induced cardiac injury. Consequently, we may have provided a new therapeutic option in the clinical prevention of DOX-induced cardiac injury. However, long-term high thyroxine levels can lead to abnormalities in cardiac structure and function,

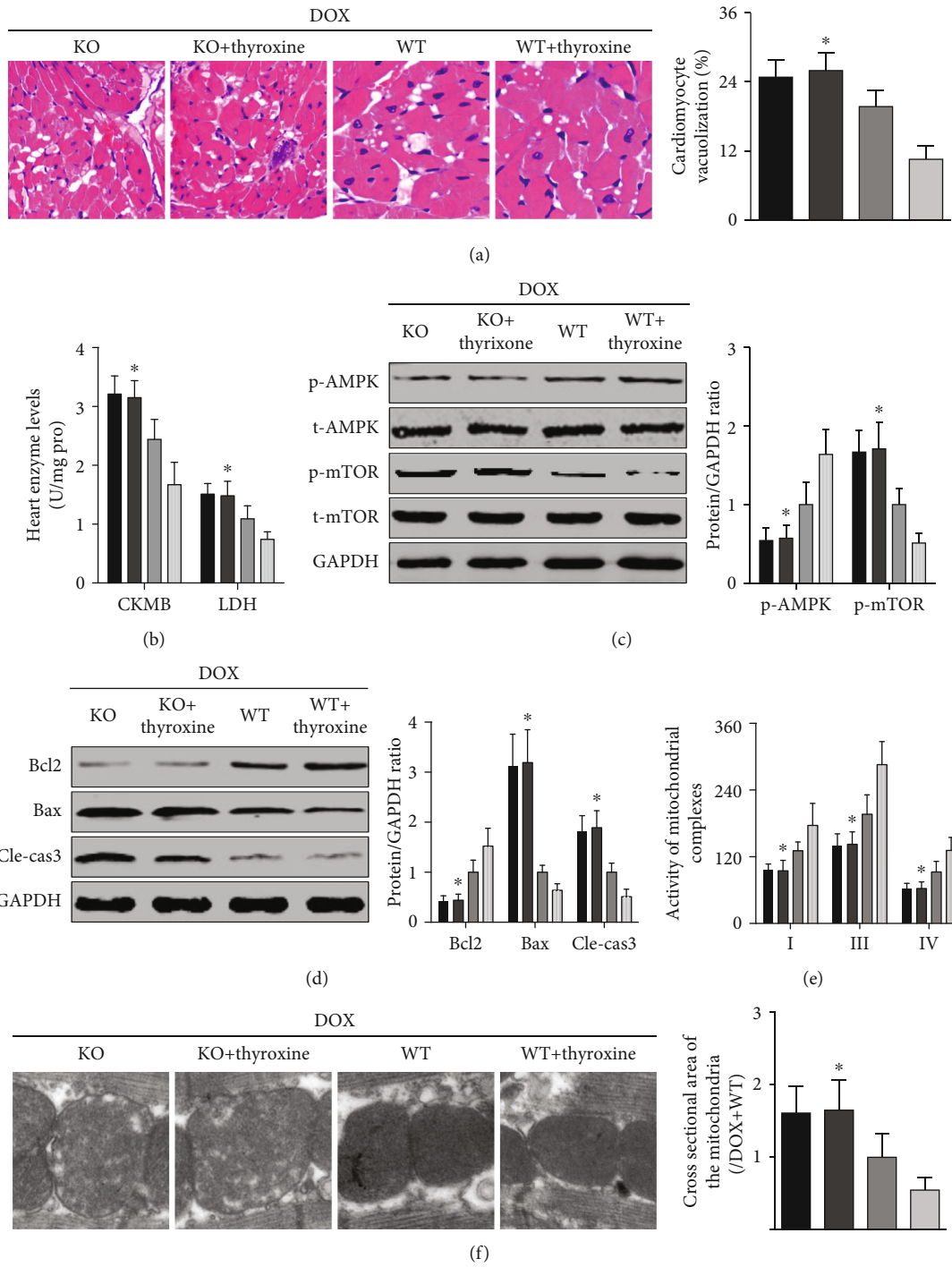


FIGURE 6: Continued.



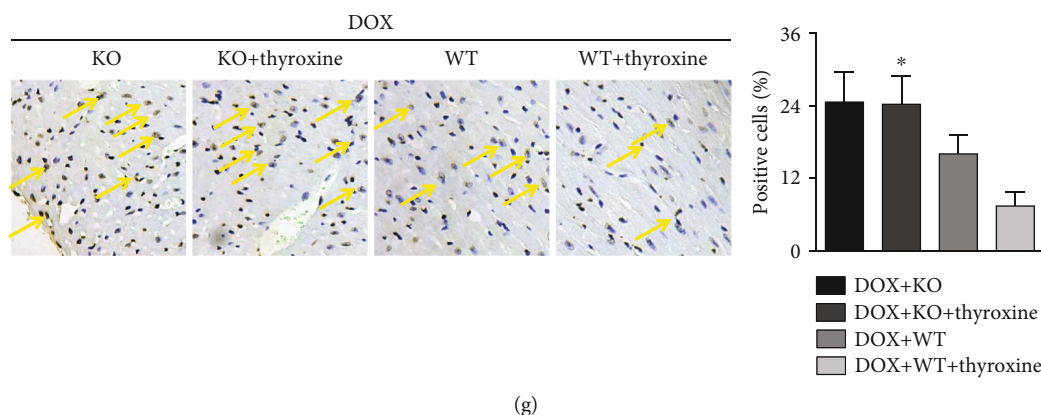


FIGURE 6: Effect of LKB1 KO on cardiac injury, energy failure, and myocardial cell apoptosis. (a) Heart sections were stained with HE, and the numbers of vacuolated cardiomyocytes in the four groups were quantified (200x). (b) Heart LDH and CK-MB levels were detected for each group. (c) Representative images and quantification of AMPK/mTOR axis phosphorylation. (d) Bcl2, Bax, and cleaved caspase-3 expression in the LV of each group. (e) Mitochondrial complex I, III, and IV activation was detected. (f) The cross-sectional area of the mitochondria in each group and more than 50 mitochondria were measured (5000x). (g) Representative images of TUNEL staining and the quantitative results for each group (200x); arrows indicate TUNEL-positive cells.  $N = 6$  in each group. \* $P < 0.05$  vs. DOX+thyroxine group.

such as those commonly observed with hyperthyroidism in the clinic. Thus, the dose and duration of thyroxine use for the prevention of DOX-induced cardiac injury need to be well controlled.

### Data Availability

We declare that the materials described in the manuscript, including all relevant raw data, will be freely available to any scientist wishing to use them for noncommercial purposes, without breaching participant confidentiality.

### Conflicts of Interest

The authors declare that they have no conflicts of interest.

### Authors' Contributions

Yuan Wang and Shan Zhu contributed equally to this work.

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