Hindawi Mediators of Inflammation Volume 2017, Article ID 9632846, 11 pages https://doi.org/10.1155/2017/9632846



Review Article

Protective Effects of Methotrexate against Proatherosclerotic Cytokines: A Review of the Evidence

Arduino A. Mangoni,¹ Angelo Zinellu,² Salvatore Sotgia,² Ciriaco Carru,^{2,3} Matteo Piga,⁴ and Gian Luca Erre⁵

Correspondence should be addressed to Arduino A. Mangoni; arduino.mangoni@flinders.edu.au

Received 20 July 2017; Revised 2 November 2017; Accepted 26 November 2017; Published 21 December 2017

Academic Editor: Sandra Helena Penha Oliveira

Copyright © 2017 Arduino A. Mangoni et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

There is good epidemiological evidence that patients with autoimmune rheumatic disease states, particularly rheumatoid arthritis, have an increased risk of cardiovascular morbidity and mortality when compared to the general population. The presence of a chronic systemic proinflammatory state in this patient group disrupts the structural and functional integrity of the endothelium and the arterial wall, favouring the onset and progression of atherosclerosis. A significant role in the detrimental effects of inflammation on endothelial function and vascular homeostasis is played by specific proatherosclerotic cytokines such as tumour necrosis factor-alpha (TNF- α), interleukin-1 (IL-1), and interleukin-6 (IL-6). Recent systematic reviews and meta-analyses have shown that treatment with methotrexate, a first-line disease-modifying antirheumatic drug (DMARD), is associated with a significant reduction in atherosclerosis-mediated cardiovascular events, such as myocardial infarction and stroke, and mortality, when compared to other DMARDs. This suggests that methotrexate might exert specific protective effects against vascular inflammation and atherosclerosis in the context of autoimmune rheumatic disease. This review discusses the available evidence regarding the potential antiatherosclerotic effects of methotrexate through the inhibition of TNF- α , IL-1, and IL-6 and provides suggestions for future experimental and human studies addressing this issue.

1. Introduction

Autoimmune rheumatic diseases such as rheumatoid arthritis (RA) are characterized by the presence of a chronic inflammatory state affecting the joints as well as a number of other organs and tissues [1]. The prevalence of RA ranges between 0.1 and 5%, depending on specific ethnic groups and geographic locations, and is higher in females than in males [1]. Patients with RA have an increased risk of death [2]. The increased mortality in this patient group, as well as in other autoimmune rheumatic conditions, is primarily due to a relatively high prevalence of cardiovascular

disease and its clinical consequences, mainly acute atherosclerosis-related events such as myocardial infarction and stroke [3–5]. The risks of myocardial infarction and stroke are, respectively, 38% and 24% higher in RA patients when compared to the general population [4, 5]. This suggests that RA favours the onset of vascular damage and atherosclerosis either through conventional cardiovascular risk factors, such as diabetes, hypercholesterolaemia, or cigarette smoking, or through alternative mechanisms [6, 7]. One possible alternative mechanism is represented by the autoimmune- and inflammation-mediated disruption of the structural and/or functional integrity of the endothelium. This

¹Department of Clinical Pharmacology, College of Medicine and Public Health, Flinders Medical Centre, Flinders University, Adelaide, SA, Australia

²Department of Biomedical Sciences, University of Sassari, Sassari, Italy

³Quality Control Unit, University Hospital of Sassari (AOUSS), Sassari, Italy

⁴Rheumatology Unit, University Clinic and AOU of Cagliari, Cagliari, Italy

⁵Rheumatology Unit, Department of Clinical and Experimental Medicine, University Hospital of Sassari (AOUSS), Sassari, Italy

leads to significant alterations in vascular homeostasis, driven by an impairment of nitric oxide (NO) synthesis by endothelial NO synthase (eNOS), that include a reduced endothelium-dependent vasodilatation, an increased leukocyte and monocyte adhesion to, and deposition in, the arterial wall, an increased intima-media thickness and arterial stiffness, and a prothrombotic tendency [8–13]. Notably, these abnormalities have been reported both in animal models of autoimmune rheumatic disease and in patients with RA [8, 14-18]. The "inflammatory theory" of atherosclerosis highlights the key role of specific cytokines, particularly tumour necrosis factor-alpha (TNF-α), interleukin-1 (IL-1), and interleukin-6 (IL-6), in disrupting endothelial integrity and vascular homeostasis [19, 20]. Given the presence of a chronic inflammatory state, and therefore a sustained vascular insult, it is likely that the pathophysiological role of proatherosclerotic cytokines is further augmented in RA. This might explain the increased cardiovascular risk reported in this patient group [7].

Over the last few years, an increasing number of in vitro and in vivo studies have sought to identify pharmacological strategies targeting inflammatory pathways for cardiovascular disease prevention and management [20-22]. A number of therapeutic agents commonly prescribed to combat immune activation and inflammation in RA might also exert salutary effects on endothelial function and vascular homeostasis. Recent epidemiological evidence suggests that methotrexate, an established first-line disease-modifying antirheumatic drug (DMARD), might exert protective effects against cardiovascular disease. This review discusses the role of cytokines and inflammation in the pathophysiology of endothelial dysfunction and atherosclerosis, the pharmacology of methotrexate, the reduced cardiovascular risk associated with its use, and the available evidence regarding the effects of this DMARD on proatherosclerotic cytokines and endothelial function.

2. Cytokines, Endothelial Dysfunction, and Atherosclerosis

The endothelium plays a key role in maintaining vascular homeostasis and protecting the vascular wall from a number of endogenous and exogenous proatherosclerotic insults [23]. A key role, in this context, is played by nitric oxide (NO), an endogenous messenger synthesised by the enzyme endothelial NO synthase (eNOS) [10]. NO regulates several important physiological processes, including vasodilation of arteries and arterioles, vascular tone, arterial stiffness, wave reflection, peripheral vascular resistance, blood pressure, and platelet function (Figure 1) [10]. A reduced NO synthesis by eNOS has been shown to be associated with virtually all cardiovascular risk factors [24]. Furthermore, clinical measures of endothelial dysfunction independently predict cardiovascular morbidity and mortality in several patient groups [25-27]. Therefore, the available evidence suggests that endothelial dysfunction is a key pathophysiological step in the sequence of events linking the presence of one or more cardiovascular risk factors with the development of atherosclerosis. Furthermore, endothelial dysfunction is useful in stratifying the risk of cardiovascular events at the population level and might represent an important target of therapies designed to mitigate such risk [28].

A significant number of cytokines, a group of lowmolecular weight proteins, are produced in several cell types, such as endothelial cells, monocytes, and vascular smooth muscle cells, that play a key role in maintaining vascular homeostasis. Amongst them, the cytokines TNF- α , IL-1, and IL-6 have been extensively investigated not only from a pathophysiological point of view but also as therapeutic targets for novel cardioprotective therapies [21, 22, 29]. Both TNF- α and IL-1 are known to stimulate the synthesis of IL-6 during the process of immune activation and inflammation [30]. This, in turn, triggers several pathways that result in endothelial dysfunction, vascular inflammation and damage, and atherosclerosis [31]. The cellular effects of TNF- α and IL-1 are primarily mediated by the p38 mitogen-activated protein kinase (p38MAPK)/nuclear factor kappa-lightchain-enhancer of the activated B-cell (NF-κB) pathways [32]. By contrast, the effects of IL-6 are mediated by the IL-6 receptor and the signal transducer protein gp130 [33].

Experimental studies have demonstrated the deleterious effects of TNF- α , IL-1, and IL-6 on endothelial function, vascular homeostasis, and cardiovascular risk and the protective effects of pharmacological agents targeting these cytokines on surrogate vascular markers (Table 1).

2.1. $TNF-\alpha$. Treatment of endothelial cells with TNF- α has been shown to increase the expression of the inducible form of NO synthase (iNOS), decreasing at the same time the expression of the endothelial constitutive isoform eNOS [34]. While the maintenance of eNOS activity provides adequate NO synthesis for the regulation of several physiological and antatherosclerotic effects, an excessive NO synthesis by iNOS leads to the intracellular formation of reactive oxygen species with consequent development of endothelial dysfunction, apoptosis, and vascular damage [35]. Not surprisingly, several studies have also reported that TNF- α significantly impairs endothelium-dependent vasodilation, through an increase in reactive oxygen species, promotes the adhesion of leukocytes to the endothelium, and favours endothelial cell apoptosis [36–38]. Furthermore, TNF- α inhibits the activity of the enzyme dimethylarginine dimethylaminohydrolase, with consequent accumulation of the endogenous eNOS inhibitor asymmetric dimethylarginine (ADMA) [39, 40]. This effect is clinically relevant as higher plasma concentrations of ADMA have been shown to independently predict cardiovascular events in patients with a wide range of cardiovascular risk at baseline (Table 1) [41, 42].

Higher serum TNF- α concentrations have been associated with an increased risk of ischaemic stroke and recurrent coronary events in epidemiological studies [43, 44]. The key role of TNF- α in mediating the detrimental effects of inflammation on endothelial dysfunction and vascular homeostasis is also supported by the results of animal and human studies investigating the effects of specific TNF- α inhibitors. For example, treatment with adalimumab significantly reduced the adhesion of human leukocytes to endothelial cells and the expression of vascular cell adhesion molecule-1

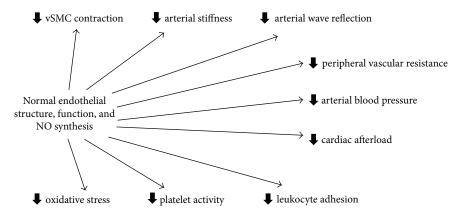


FIGURE 1: Endothelium, nitric oxide, and vascular homeostasis. NO: nitric oxide; VSMC: vascular smooth muscle cell.

Table 1: Effects of the cytokines TNF-α, IL-1, and IL-6 on endothelial function and vascular homeostasis.

Cytokine	Reported effects
Tumour necrosis factor-alpha	Endothelial nitric oxide synthase activity \
	Inducible nitric oxide synthase activity ↑
	Reactive oxygen species ↑
	Endothelium-dependent vasodilation \downarrow
	Leukocyte adhesion ↑
	Endothelial cell apoptosis ↑
	Asymmetric dimethylarginine ↑
Interleukin-1	Endothelium-dependent vasodilation ↓
	Vasoconstrictor response to
	pharmacological challenge ↑
	Endothelin-1 ↑
	Leukocyte adhesion ↑
	Vascular smooth muscle cell growth ↑
	Intima thickness ↑
	Arterial stiffness ↑
	Coagulation ↑
Interleukin-6	Expression of angiotensin II type-1
	receptor ↑
	Endothelium-dependent vasodilation \downarrow
	Arterial stiffness ↑
	Oxidative stress ↑
	Thrombosis ↑

↑: Increase; ↓: decrease.

(VCAM-1), intracellular adhesion molecule-1 (ICAM-1), and E-selectin [45]. Other studies have shown that treatment with adalimumab or etanercept improved clinical measures of endothelium-dependent vasodilation in patients with psoriasis, reduced the plasma concentrations of ADMA, and increased circulating endothelial progenitor cells in patients with RA (Table 1) [46–48].

2.2. IL-1. Increased production of IL-1 has been shown to induce leukocyte adhesion to the endothelium, exert procoagulant activity, and stimulate the growth and chemotaxis of vascular smooth muscle cells, key steps in the pathogenesis

of atherosclerosis [49–52]. In animal studies, exposure to exogenous IL-6 causes an increase in coronary vasospastic responses to pharmacological challenge and intima thickening (Table 1) [53]. Moreover, multiple factors known to associate with atherosclerosis, such as cholesterol crystals, atheroprone oscillatory flow, hypoxia, and neutrophil extracellular traps, have recently been found to activate the critical IL-1beta producing NLRP3 inflammasome [22].

Polymorphisms in the IL-1 receptor antagonist (IL-1Ra) gene have been shown to have significant associations with the presence of single-vessel coronary artery disease, assessed by coronary angiography, in patients with ischaemic heart disease [54]. Conversely, pharmacological inhibition of IL-1 with anakinra reduces the concentrations of the potent endogenous vasoconstrictor endothelin-1 and arterial stiffness and improves clinical measures of endothelial function in patients with RA [55]. Similar effects of anakinra on endothelium-dependent vasodilation have been reported in animal models of diabetes (Table 1) [56]. In a recently completed randomized placebo-controlled trial in 10,061 participants with a previous myocardial infarction and C-reactive protein concentrations ≥2 mg/L, canakinumab, a monoclonal antibody targeting IL-1beta, significantly reduced the primary end-point of nonfatal myocardial infarction, nonfatal stroke, or cardiovascular death after a median follow-up of 3.7 years. Of the three subcutaneous doses of canakinumab studied (50 mg, 150 mg, and 300 mg every three months), only the 150 mg dose met the prespecified multiplicity adjusted threshold for statistical significance. Furthermore, there were no significant differences in all-cause mortality between treatment with canakinumab and placebo [57].

2.3. IL-6. IL-6 has been shown to upregulate the expression of the angiotensin II type-1 receptor in vascular smooth muscle cells, responsible for vasoconstriction, cell apoptosis, and proinflammatory effects, and to impair endothelium-dependent vasodilation in animal models [58]. Increased plasma IL-6 concentrations are significantly associated with a reduced endothelium-dependent vasodilation both in healthy subjects [59] and in patients with hypertension [60] or hypercholesterolaemia [61]. A significant association between plasma IL-6 concentrations and clinical markers of increased arterial stiffness has been reported in patients with

hypertension (Table 1) [62]. A meta-analysis performed by the Emerging Risk Factors Collaboration has also shown that higher plasma IL-6 concentrations predict the risk of nonfatal myocardial infarction and coronary artery disease-related death, with a 25% increase in the risk of future vascular events for each increase in log IL-6 concentrations (RR 1.25, 95% CI 1.19 to 1.32) [63].

Furthermore, treatment with the IL-6 receptor inhibitor tocilizumab caused an increase in endothelium-dependent vasodilatation, and a concomitant reduction in arterial stiffness, in patients with RA [64, 65]. In another study, tocilizumab improved endothelial function and reduced markers of oxidative stress, inflammation, and thrombosis in patients with RA (Table 1) [66].

3. Methotrexate Pharmacology

Methotrexate, an analogue of the B vitamin folic acid, is a first-line synthetic DMARD for the management of RA and other autoimmune diseases that is normally administered once a week, either orally, subcutaneously, or intramuscularly, with doses ranging between 5 and 25 mg [67, 68]. Notably, methotrexate is the only DMARD that has demonstrated significant survival benefits in patients with RA [69-71]. After being transported into the cytoplasm, through the reduced folate carrier, methotrexate is converted into intracellular polyglutamates. The polyglutamate forms ensure the intracellular retention of methotrexate, allowing weekly administration despite the relatively short plasma elimination half-life (5-8 hours) [72]. The polyglutamates also mediate the immunomodulating and anti-inflammatory effects of methotrexate, by reducing the synthesis of purines, pyrimidines, and DNA through the inhibition of dihydrofolate reductase, thymidylate synthase, and aminoimidazole carboxamide ribonucleotide (AICAR) transformylase (ATIC, Figure 2) [73–75]. Furthermore, the inhibition of ATIC causes the accumulation of the substrate AICAR which, in turn, inhibits the enzymes adenosine deaminase and adenosine monophosphate (AMP) deaminase, involved in the catabolism of adenosine (Figure 2) [75]. Adenosine per se exerts significant anti-inflammatory effects, primarily through the A_{2A} and A₃ receptors [76], and mediates the anti-inflammatory effects of methotrexate. In animal models of inflammation, methotrexate has been shown to increase AICAR and adenosine concentrations and to inhibit the accumulation of leukocytes, in exudates from carrageenaninflamed air pouches. The administration of AMP deaminase partly reversed the methotrexate-mediated reduction in leukocyte accumulation. Additionally, the administration of 3,7-dimethyl-1-propargylxanthine, an A2A receptor antagonist, but not 8-cyclopentyl-dipropylxanthine, an A₁ receptor antagonist, suppressed the methotrexate-mediated reduction in leukocyte accumulation [77]. Furthermore, both AICAR and AMP activate the 5' adenosine monophosphateactivated protein kinase (AMPK) [78]. AMPK activation is protective towards endothelial cell function and vascular homeostasis, by ensuring physiological NO synthesis, maintaining mitochondrial structure and function, and preventing oxidative stress and apoptosis [79]. Moreover, a role for

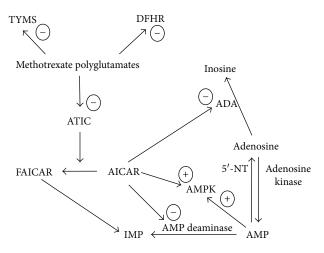


FIGURE 2: Intracellular effects of methotrexate. DHFR: dihydrofolate reductase; TYMS: thymidylate synthase; ATIC: aminoimidazole carboxamide ribonucleotide (AICAR) transformylase; FAICAR: 5-formamidoimidazole-4-carboxamide ribotide; IMP: inosine monophosphate; AMP: adenosine monophosphate; AMPK: 5' adenosine monophosphate-activated protein kinase; ADA: adenosine deaminase; 5'-NT: 5'-nucleotidase; -: inhibition; +: activation

AMPK in inhibiting vascular smooth muscle cell proliferation, a key proatherosclerotic event both in autoimmune rheumatic diseases and in the general population, has been proposed [80, 81]. There is experimental evidence that AMPK is a key mediator of the effects of methotrexate on inflammation and endothelial function. In vitro studies have shown that methotrexate, at concentrations between 0.1 and 0.5 µM, increased AMPK phosphorylation and AMPK activity in perivascular adipose tissue cells. In these cells, the administration of palmitic acid, a proinflammatory agent, decreased AMPK phosphorylation; however, these effects were significantly reduced by pretreatment with either methotrexate or AICAR. Furthermore, methotrexate significantly reduced the phosphorylation of NF-κB p65 and prevented the palmitic acid-mediated increase in the expression of TNF- α and IL-6, indicating anti-inflammatory effects. Silencing AMPK with AMPKa1/2-specific siRNA significantly decreased the inhibitory effects of methotrexate on palmitic acid-mediated NF-κB p65 phosphorylation and on the expression of TNF- α and IL-6. Pretreatment with methotrexate prevented the impairment of acetylcholine-induced, endothelium-dependent vasodilation in rat aorta following palmitic acid administration. Notably, the protective effects of methotrexate on endothelial function were attenuated by cotreatment with compound C, an AMPK inhibitor [82].

4. Methotrexate and Cardiovascular Risk

Recent systematic reviews and meta-analyses have investigated the associations between methotrexate treatment and cardiovascular risk. Micha et al. identified observational studies in 66,334 patients with either RA (9 studies), psoriasis (1 study), or polyarthritis (1 study), that reported 6235 cardiovascular events. The median duration of follow-up in

these studies was 5.8 years. Six studies compared methotrexate users versus never users, three compared current versus noncurrent users, and two compared initiators versus noninitiators. Cardiovascular endpoints included total or fatal cardiovascular disease (7 studies; two studies also provided separate estimates for myocardial infarction and ischaemic/haemorrhagic stroke), myocardial infarction (3 studies), and ischaemic stroke (1 study). Combined assessment of the included studies showed that methotrexate treatment was associated with a significant reduction in cardiovascular events (RR 0.79, 95% CI 0.73 to 0.87). Assessment of specific cardiovascular endpoints showed a similar effect size (overall cardiovascular disease events: RR 0.76, 95% CI 0.69 to 0.84; myocardial infarction: RR 0.82, 95% CI 0.71 to 0.96; stroke: RR 0.70, 95% CI 0.56 to 0.87) [83]. In a subsequent systematic review and meta-analysis, Roubille et al. identified 34 studies (28 studies in 236,525 RA patients, reporting 5410 cardiovascular events, and 6 studies in 220,209 patients with either psoriasis or psoriatic arthritis, reporting 2701 cardiovascular events). In studies conducted in RA patients, methotrexate use was associated with a reduced risk of all cardiovascular events (RR 0.72, 95% CI 0.57 to 0.91, P = 0.007) and myocardial infarction (RR 0.81, 95% CI 0.68 to 0.86, P = 0.01) when compared to other synthetic DMARDs. Although there was no significant effect of methotrexate on the risk of either stroke (RR 0.78, 95% CI 0.40 to 1.50) or major adverse cardiovascular events (RR 0.38, 95% CI 0.05 to 2.84), the number of studies assessing these endpoints was relatively low (one study for stroke and two studies for major adverse cardiovascular events, resp.) [84]. Therefore, the results of observational studies support the hypothesis that methotrexate exhibits specific protective cardiovascular effects when compared to other DMARDs.

5. Methotrexate and Proatherosclerotic Cytokines

In order to review the available evidence on the direct or indirect, mediated by adenosine, AICAR or AMPK activation, effects of methotrexate on proatherosclerotic cytokines, a PubMed literature search was conducted from inception to June 2017, using the following terms: methotrexate, adenosine, AICAR, AMPK, endothelium, inflammation, atherosclerosis, TNF- α , IL-1, and IL-6.

5.1. $TNF-\alpha$. In *in vitro* studies, clinical concentrations of methotrexate $(2\times10^{-8}\,\mathrm{M}$ and $2\times10^{-7}\,\mathrm{M})$ have been shown to increase the release of soluble TNF receptor p75 from the cell surface [85]. This phenomenon is able to significantly inhibit the proinflammatory effects of TNF- α [86]. Adenosine is a known inhibitor of TNF- α expression through stimulation of the A_3 receptor (Table 2) [87, 88]. There is recent evidence that methotrexate stimulates AMPK phosphorylation and activity, induces manganese superoxide dismutase mRNA and protein, and increases the expression of the cytoprotective genes haem oxygenase-1 and Bcl-2-related protein in human umbilical vein endothelial cells (HUVECs) and arterial endothelial cells (HAECs). The pretreatment of endothelial cells with TNF- α did not affect

Table 2: Effects of methotrexate, adenosine, AICAR, and AMPK activation on endothelial function and vascular homeostasis.

Mediator	Reported effects
Methotrexate	Release of soluble TNF-α receptor p75 ↑
	TNF- α expression/concentrations \downarrow
	IL-6 expression/concentrations ↓
	ICAM-1 expression ↓
	VCAM-1 expression ↓
	eNOS activity ↑
	Endothelium-dependent vasodilatation \uparrow
	Mitochondrial mass, membrane potential, and intracellular ATP concentrations ↑
Adenosine	TNF- α expression/concentrations \downarrow
	IL-6 expression/concentrations ↓
	ICAM-1 expression ↓
	VCAM-1 expression ↓
	E-selectin expression \downarrow
	eNOS activity ↑
	Blood pressure ↓
	Mitochondrial mass, membrane potential, and intracellular ATP concentrations ↑
	Formation of atherosclerotic lesions \downarrow
	Cholesterol concentrations \downarrow
	Triglyceride concentrations \downarrow
	IL-1 expression/concentrations \downarrow
AICAR/AMPK	IL-6 expression/concentrations \downarrow
	ICAM-1 expression ↓
	VCAM-1 expression ↓
	NO synthesis ↑
	Endothelium-dependent vasodilation \uparrow
	Endothelium-independent vasodilation \uparrow
	Blood pressure ↓
	Oxidative stress ↓
	Endoplasmic reticulum stress ↓
	Manganese superoxide dismutase induction 1
	NF-κB ↓
	Monocyte adhesion to endothelial cells \downarrow
	Restenosis ↓
	Cholesterol efflux capacity ↑
	Cellular glucose uptake ↑
	Glycolysis ↑

AICAR: aminoimidazole carboxamide ribonucleotide; AMPK: 5' adenosine monophosphate-activated protein kinase; TNF- α : tumour necrosis factoralpha; IL-1: interleukin-1; IL-6: interleukin-6; ICAM-1: intercellular adhesion molecule-1; VCAM-1: vascular cell adhesion molecule-1; ATP: adenosine triphosphate; eNOS: endothelial nitric oxide synthase; NF- α B: nuclear factor kappa-light-chain-enhancer of the activated B-cell; NO: nitric oxide; \uparrow : increase; \downarrow : decrease.

the methotrexate-mediated upregulation of manganese superoxide dismutase and haem oxygenase-1 mRNA [89]. Methotrexate (10^{-8} M to 10^{-6} M) and/or adenosine treatment has also been shown to prevent the TNF- α -induced (a) expression of ICAM-1 and VCAM-1 in human

umbilical vein endothelial cells (HUVEC) [90] and (b) reduction of mitochondrial mass, membrane potential, and intracellular ATP concentrations, in endothelial cells through increased activity of eNOS [91]. Furthermore, adenosine has been shown to reduce the circulating concentrations of TNF- α and IL-6 in a mice model of sepsis (Table 2) [92]. By contrast, therapeutic concentrations of methotrexate, 0.1, 0.25, and 0.5 μ M, significantly increased the expression of the TNF- α receptor-associated factor 1 gene (~threefold change) and of the TNF- α receptor superfamily member 9 gene (~threefold change) in EA.hy 926 cells, derived from the fusion of primary endothelial cells with an epithelial tumour cell line [93].

5.2. IL-1. In an *in vitro* model of palmitate-induced endothelial dysfunction, AICAR-mediated AMPK activation has been shown to significantly reduce IL-1, IL-6, and VCAM-1 synthesis, by preventing the activation of the NLRP3 inflammasome [94]. Similarly, pharmacological activation of AMPK with oestradiol prevented the IL-1-induced expression of VCAM-1 and ICAM-1 in cultured human endothelial cells (Table 2) [95]. By contrast, therapeutic concentrations of methotrexate, 0.1, 0.25, and 0.5 μ M, significantly increased the expression of the IL-1 alpha gene (~threefold change) and of the IL-1 receptor-like 1 gene (~fourfold change) in EA.hy 926 cells [93].

5.3. IL-6. In db/db mice fed with Western diet, methotrexate treatment (4 mg/kg) caused a significant reduction in the circulating concentrations of IL-6 and TNF- α . These effects were associated with a significant increase in endotheliumdependent vasodilatation and a reduction in VCAM-1 [96]. In another study, methotrexate treatment $(0.1-0.5 \,\mu\text{M})$ significantly reduced the expression of IL-6 and TNF- α . These effects were mediated by the activation of AMPK and were associated with an improvement in eNOS activity and endothelium-dependent vasodilation in rat aorta (Table 2) [82]. Adenosine has been shown to prevent the thrombinmediated increased expression of IL-6, VCAM-1, ICAM-1, and E-selectin in HUVECs. These effects are mediated by the activation of the adenosine receptor A_{2A} [97]. Similarly, adenosine inhibited, in a dose-dependent fashion, the release of IL-6, VCAM-1, and ICAM-1 in HUVECs pretreated with either IL-1, TNF- α , or lipopolysaccharide (Table 2) [98].

6. Additional Vascular Effects of Adenosine, AICAR, and AMPK Activation

A number of studies have shown that adenosine, AICAR, and AMPK activation provide beneficial effects on endothelial function and vascular homeostasis that are independent of those on TNF- α , IL-1, or IL-6.

6.1. Adenosine Accumulation. There is good evidence that adenosine, through the activation of the A_{2A} and A_{2B} receptors, lowers blood pressure as a result of increased NO synthesis in the endothelium and direct vasodilation [99, 100]. Studies have also demonstrated the presence of central nervous system-mediated hypotensive effects through the

activation of the A_3 receptor [101]. Furthermore, the pharmacological activation of the A_{2B} receptor prevents the formation of atherosclerotic lesions and reduces the plasma concentrations of cholesterol and triglycerides, possibly through the reduced activation of the transcription factor sterol regulatory element-binding protein 1 in the liver [102, 103]. Activation of adenosine receptors seems also to modulate glucose homeostasis. However, the pathophysiological and clinical relevance of this finding is yet to be determined (Table 2) [104].

6.2. AICAR and Activation of 5' Adenosine Monophosphate-Activated Protein Kinase (AMPK). There is increasing evidence that AICAR and or/AMPK activation stimulates NO synthesis in the endothelium, enhances endotheliumdependent and endothelium-independent vasodilation, reduces blood pressure, prevents vessel restenosis, and increases cholesterol efflux capacity [105–110]. Furthermore, AMPK stimulates cellular glucose uptake, through GLUT-1 and GLUT-4 transporters, and glycolysis, through phosphorylation of two isoforms of the enzyme 6-phoshofructo-2kinase: fructose-2,6-biphosphatase, with beneficial effects on glucose homeostasis [111-114]. There is also evidence that AMPK activation protects endothelial cells from the deleterious effects of chronic exposure to high concentrations of glucose and fatty acids and consequently reduces oxidative stress, inflammation, and endoplasmic reticulum stress (Table 2) [115, 116].

7. Discussion

The available evidence from systematic reviews and metaanalyses of observational studies in patients with either RA or other autoimmune rheumatic disease states suggests that the use of methotrexate is associated with a significant reduction in cardiovascular morbidity and mortality when compared to other DMARDs. Furthermore, a relatively small number of experimental studies have shown that methotrexate can exert beneficial effects on endothelial function and vascular homeostasis by preventing or blocking the effects of key proatherosclerotic cytokines such as TNF- α , IL-1, and IL-6. The reported effects are mediated either by methotrexate directly or through the activation of adenosine receptors, AICAR, or AMPK (Table 2). Pending further in vitro and in vivo studies investigating the exact mechanisms involved in such effects, a number of limitations need to be considered when interpreting the available data:

- (1) The dose of methotrexate used in some studies [96] and its consequent local concentrations in target cells and tissues are quite different from those normally observed in patients with autoimmune disorders.
- (2) No study has assessed the intracellular concentrations of methotrexate polyglutamates, as a factor mediating the effects of the drug on the study end-points.
- (3) The effects of methotrexate, adenosine, AICAR, or AMPK activation on proatherosclerotic cytokines

were not compared to those of other synthetic or biologic DMARDs.

- (4) Other studies, not specifically investigating TNF-α, IL-1, and IL-6, have shown that methotrexate can also exert antiproliferative effects in human umbilical vein endothelial cells and EA.hy 926 cells [93, 117, 118] and reduce NO synthesis, possibly as a result of reduced availability of tetrahydrobiopterin, an essential cofactor for endothelial nitric oxide synthase, secondary to dihydrofolate reductase inhibition [119].
- (5) There is no direct evidence that the methotrexateinduced inhibition of TNF- α , IL-1, or IL-6 leads to sustained beneficial effects on endothelial function, atherosclerosis, arterial structure and function, and cardiovascular risk in human studies.

These issues should be accounted for in future studies investigating the effects of methotrexate on proatherosclerotic cytokines. In particular, the use of other DMARDs as comparator should help to determine whether methotrexate treatment exerts specific antiatherosclerotic effects that might help to explain its superiority, in terms of cardiovascular risk reduction, reported in observational studies. Furthermore, a comprehensive assessment of proatherosclerotic and antiatherosclerotic cytokines, such as transforming growth factor- β , IL-10, and IL-35 [120], as well as measures of endothelial cell proliferation, apoptosis, and nitric oxide synthesis, might provide additional mechanistic insights regarding the possible vasculoprotective effects of methotrexate and the potential rationale for combining methotrexate treatment with other interventional strategies targeting specific cytokines. In this context, the results of the Cardiovascular Inflammation Reduction Trial (CIRT, clinicaltrials.gov identifier NCT01594333), investigating the effects of methotrexate on myocardial infarction, stroke, and cardiovascular death in patients with type 2 diabetes or metabolic syndrome and stable coronary artery disease, will provide additional knowledge regarding the potential repurposing of methotrexate for cardiovascular risk management and prevention in different patient populations [121].

8. Conclusions

The evidence recently generated from experimental studies suggests that methotrexate, an anchor drug in the treatment of RA and other autoimmune disorders, might exert significant beneficial effects on endothelial function and vascular homeostasis by targeting key cytokines regulating the immune responses and inflammation pathways responsible for the development of atherosclerosis and thrombosis. However, further studies are required to fully establish the role of this DMARD as an antiatherosclerotic and cardioprotective agent.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

References

- [1] J. S. Smolen, D. Aletaha, and I. B. McInnes, "Rheumatoid arthritis," *The Lancet*, vol. 388, no. 10055, pp. 2023–2038, 2016.
- [2] A. Gonzalez, H. Maradit Kremers, C. S. Crowson et al., "The widening mortality gap between rheumatoid arthritis patients and the general population," *Arthritis & Rheumatism*, vol. 56, no. 11, pp. 3583–3587, 2007.
- [3] M. Piga, L. Casula, D. Perra et al., "Population-based analysis of hospitalizations in a West-European region revealed major changes in hospital utilization for patients with systemic lupus erythematosus over the period 2001-2012," *Lupus*, vol. 25, no. 1, pp. 28–37, 2016.
- [4] W. S. Chung, C. L. Lin, C. L. Peng et al., "Rheumatoid arthritis and risk of acute myocardial infarction—a nationwide retrospective cohort study," *International Journal of Cardiology*, vol. 168, no. 5, pp. 4750–4754, 2013.
- [5] T. H. Liou, S. W. Huang, J. W. Lin, Y. S. Chang, C. W. Wu, and H. W. Lin, "Risk of stroke in patients with rheumatism: a nationwide longitudinal population-based study," *Scientific Reports*, vol. 4, p. 5110, 2014.
- [6] L. R. Baghdadi, R. J. Woodman, E. M. Shanahan, and A. A. Mangoni, "The impact of traditional cardiovascular risk factors on cardiovascular outcomes in patients with rheumatoid arthritis: a systematic review and meta-analysis," *PLoS One*, vol. 10, no. 2, article e0117952, 2015.
- [7] K. Lauper and C. Gabay, "Cardiovascular risk in patients with rheumatoid arthritis," *Seminars in Immunopathology*, vol. 39, no. 4, pp. 447–459, 2017.
- [8] G. Murdaca, B. M. Colombo, P. Cagnati, R. Gulli, F. Spano, and F. Puppo, "Endothelial dysfunction in rheumatic autoimmune diseases," *Atherosclerosis*, vol. 224, no. 2, pp. 309–317, 2012.
- [9] S. Cardaropoli, F. Silvagno, E. Morra, G. P. Pescarmona, and T. Todros, "Infectious and inflammatory stimuli decrease endothelial nitric oxide synthase activity *in vitro*," *Journal of Hypertension*, vol. 21, no. 11, pp. 2103–2110, 2003.
- [10] C. Napoli, F. de Nigris, S. Williams-Ignarro, O. Pignalosa, V. Sica, and L. J. Ignarro, "Nitric oxide and atherosclerosis: an update," *Nitric Oxide*, vol. 15, no. 4, pp. 265–279, 2006.
- [11] J. Loscalzo, "Nitric oxide insufficiency, platelet activation, and arterial thrombosis," *Circulation Research*, vol. 88, no. 8, pp. 756–762, 2001.
- [12] R. D. Rudic and W. C. Sessa, "Nitric oxide in endothelial dysfunction and vascular remodeling: clinical correlates and experimental links," *American Journal of Human Genetics*, vol. 64, no. 3, pp. 673–677, 1999.
- [13] I. B. Wilkinson, S. S. Franklin, and J. R. Cockcroft, "Nitric oxide and the regulation of large artery stiffness: from physiology to pharmacology," *Hypertension*, vol. 44, no. 2, pp. 112–116, 2004.
- [14] Y. Haruna, Y. Morita, N. Komai et al., "Endothelial dysfunction in rat adjuvant-induced arthritis: vascular superoxide production by NAD(P)H oxidase and uncoupled endothelial nitric oxide synthase," *Arthritis & Rheumatism*, vol. 54, no. 6, pp. 1847–1855, 2006.
- [15] S. E. Abbot, W. J. Whish, C. Jennison, D. R. Blake, and C. R. Stevens, "Tumour necrosis factor α stimulated rheumatoid synovial microvascular endothelial cells exhibit increased shear rate dependent leucocyte adhesion in vitro," *Annals of the Rheumatic Diseases*, vol. 58, no. 9, pp. 573–581, 1999.

[16] P. Wang, S. Y. Guan, S. Z. Xu et al., "Increased carotid intima-media thickness in rheumatoid arthritis: an update meta-analysis," *Clinical Rheumatology*, vol. 35, no. 2, pp. 315–323, 2016.

- [17] M. J. Roman, R. B. Devereux, J. E. Schwartz et al., "Arterial stiffness in chronic inflammatory diseases," *Hypertension*, vol. 46, no. 1, pp. 194–199, 2005.
- [18] J. Beinsberger, J. W. Heemskerk, and J. M. Cosemans, "Chronic arthritis and cardiovascular disease: altered blood parameters give rise to a prothrombotic propensity," *Seminars in Arthritis & Rheumatism*, vol. 44, no. 3, pp. 345–352, 2014.
- [19] P. Libby, "Inflammation in atherosclerosis," Arteriosclerosis, Thrombosis, and Vascular Biology, vol. 32, no. 9, pp. 2045– 2051, 2012.
- [20] D. Tousoulis, E. Oikonomou, E. K. Economou, F. Crea, and J. C. Kaski, "Inflammatory cytokines in atherosclerosis: current therapeutic approaches," *European Heart Journal*, vol. 37, no. 22, pp. 1723–1732, 2016.
- [21] D. P. Ramji and T. S. Davies, "Cytokines in atherosclerosis: key players in all stages of disease and promising therapeutic targets," *Cytokine & Growth Factor Reviews*, vol. 26, no. 6, pp. 673–685, 2015.
- [22] P. M. Ridker, "From C-reactive protein to interleukin-6 to interleukin-1: moving upstream to identify novel targets for atheroprotection," *Circulation Research*, vol. 118, no. 1, pp. 145–156, 2016.
- [23] C. M. Boulanger, "Endothelium," *Arteriosclerosis, Thrombosis, and Vascular Biology*, vol. 36, no. 4, pp. e26–e31, 2016.
- [24] M. K. Reriani, L. O. Lerman, and A. Lerman, "Endothelial function as a functional expression of cardiovascular risk factors," *Biomarkers in Medicine*, vol. 4, no. 3, pp. 351–360, 2010.
- [25] Y. Matsuzawa, T. G. Kwon, R. J. Lennon, L. O. Lerman, and A. Lerman, "Prognostic value of flow-mediated vasodilation in brachial artery and fingertip artery for cardiovascular events: a systematic review and meta-analysis," *Journal of the American Heart Association*, vol. 4, no. 11, article e002270, 2015.
- [26] Y. Xu, R. C. Arora, B. M. Hiebert et al., "Non-invasive endothelial function testing and the risk of adverse outcomes: a systematic review and meta-analysis," *European Heart Journal Cardiovascular Imaging*, vol. 15, no. 7, pp. 736–746, 2014.
- [27] R. T. Ras, M. T. Streppel, R. Draijer, and P. L. Zock, "Flow-mediated dilation and cardiovascular risk prediction: a systematic review with meta-analysis," *International Journal of Cardiology*, vol. 168, no. 1, pp. 344–351, 2013.
- [28] H. A. Jensen and J. L. Mehta, "Endothelial cell dysfunction as a novel therapeutic target in atherosclerosis," *Expert Review of Cardiovascular Therapy*, vol. 14, no. 9, pp. 1021–1033, 2016.
- [29] A. Tedgui and Z. Mallat, "Cytokines in atherosclerosis: pathogenic and regulatory pathways," *Physiological Reviews*, vol. 86, no. 2, pp. 515–581, 2006.
- [30] M. Feldmann, F. M. Brennan, and R. N. Maini, "Role of cytokines in rheumatoid arthritis," *Annual Review of Immu*nology, vol. 14, no. 1, pp. 397–440, 1996.
- [31] P. Libby and P. M. Ridker, "Novel inflammatory markers of coronary risk: theory versus practice," *Circulation*, vol. 100, no. 11, pp. 1148–1150, 1999.
- [32] K. F. Chan, M. R. Siegel, and J. M. Lenardo, "Signaling by the TNF receptor superfamily and T cell homeostasis," *Immunity*, vol. 13, no. 4, pp. 419–422, 2000.

[33] H. Ait-Oufella, S. Taleb, Z. Mallat, and A. Tedgui, "Recent advances on the role of cytokines in atherosclerosis," *Arteriosclerosis, Thrombosis, and Vascular Biology*, vol. 31, no. 5, pp. 969–979, 2011.

- [34] K. L. MacNaul and N. I. Hutchinson, "Differential expression of iNOS and cNOS mRNA in human vascular smooth muscle cells and endothelial cells under normal and inflammatory conditions," *Biochemical and Biophysical Research Commu*nications, vol. 196, no. 3, pp. 1330–1334, 1993.
- [35] W. N. Nowak, J. Deng, X. Z. Ruan, and Q. Xu, "Reactive oxygen species generation and atherosclerosis," *Arterioscle*rosis, Thrombosis, and Vascular Biology, vol. 37, no. 5, pp. e41–e52, 2017.
- [36] X. Gao, S. Belmadani, A. Picchi et al., "Tumor necrosis factor-α induces endothelial dysfunction in Lepr^{db} mice," *Circulation*, vol. 115, no. 2, pp. 245–254, 2007.
- [37] A. Picchi, X. Gao, S. Belmadani et al., "Tumor necrosis factor-α induces endothelial dysfunction in the prediabetic metabolic syndrome," *Circulation Research*, vol. 99, no. 1, pp. 69–77, 2006.
- [38] L. A. Madge and J. S. Pober, "TNF signaling in vascular endothelial cells," *Experimental and Molecular Pathology*, vol. 70, no. 3, pp. 317–325, 2001.
- [39] A. Ito, P. S. Tsao, S. Adimoolam, M. Kimoto, T. Ogawa, and J. P. Cooke, "Novel mechanism for endothelial dysfunction: dysregulation of dimethylarginine dimethylaminohydrolase," *Circulation*, vol. 99, no. 24, pp. 3092–3095, 1999.
- [40] C. Wadham and A. A. Mangoni, "Dimethylarginine dimethylaminohydrolase regulation: a novel therapeutic target in cardiovascular disease," *Expert Opinion on Drug Metabolism & Toxicology*, vol. 5, no. 3, pp. 303–319, 2009.
- [41] P. Willeit, D. F. Freitag, J. A. Laukkanen et al., "Asymmetric dimethylarginine and cardiovascular risk: systematic review and meta-analysis of 22 prospective studies," *Journal of the American Heart Association*, vol. 4, no. 6, article e001833, 2015.
- [42] C. Xuan, Q. W. Tian, H. Li, B. B. Zhang, G. W. He, and L. M. Lun, "Levels of asymmetric dimethylarginine (ADMA), an endogenous nitric oxide synthase inhibitor, and risk of coronary artery disease: a meta-analysis based on 4713 participants," *European Journal of Preventive Cardiology*, vol. 23, no. 5, pp. 502–510, 2016.
- [43] G. Cui, H. Wang, R. Li et al., "Polymorphism of tumor necrosis factor alpha (TNF-alpha) gene promoter, circulating TNF-alpha level, and cardiovascular risk factor for ischemic stroke," *Journal of Neuroinflammation*, vol. 9, p. 235, 2012.
- [44] P. M. Ridker, N. Rifai, M. Pfeffer, F. Sacks, S. Lepage, and E. Braunwald, "Elevation of tumor necrosis factor-α and increased risk of recurrent coronary events after myocardial infarction," *Circulation*, vol. 101, no. 18, pp. 2149–2153, 2000
- [45] C. Rios-Navarro, C. de Pablo, V. Collado-Diaz et al., "Differential effects of anti-TNF- α and anti-IL-12/23 agents on human leukocyte-endothelial cell interactions," *European Journal of Pharmacology*, vol. 765, pp. 355–365, 2015.
- [46] G. Avgerinou, D. Tousoulis, G. Siasos et al., "Anti-tumor necrosis factor alpha treatment with adalimumab improves significantly endothelial function and decreases inflammatory process in patients with chronic psoriasis," *Interna*tional Journal of Cardiology, vol. 151, no. 3, pp. 382-383, 2011.

[47] F. R. Spinelli, M. Di Franco, A. Metere et al., "Decrease of asymmetric dimethyl arginine after anti-TNF therapy in patients with rheumatoid arthritis," *Drug Development Research*, vol. 75, Supplement 1, pp. S67–S69, 2014.

- [48] F. R. Spinelli, A. Metere, C. Barbati et al., "Effect of therapeutic inhibition of TNF on circulating endothelial progenitor cells in patients with rheumatoid arthritis," *Mediators of Inflammation*, vol. 2013, Article ID 537539, 8 pages, 2013.
- [49] M. P. Bevilacqua, J. S. Pober, M. E. Wheeler, R. S. Cotran, and M. A. Gimbrone Jr., "Interleukin 1 acts on cultured human vascular endothelium to increase the adhesion of polymorphonuclear leukocytes, monocytes, and related leukocyte cell lines," *The Journal of Clinical Investigation*, vol. 76, no. 5, pp. 2003–2011, 1985.
- [50] M. P. Bevilacqua, J. S. Pober, G. R. Majeau, R. S. Cotran, and M. A. Gimbrone Jr, "Interleukin 1 (IL-1) induces biosynthesis and cell surface expression of procoagulant activity in human vascular endothelial cells," *The Journal of Experimental Medicine*, vol. 160, no. 2, pp. 618–623, 1984.
- [51] U. Ikeda, M. Ikeda, T. Oohara, S. Kano, and T. Yaginuma, "Mitogenic action of interleukin- 1α on vascular smooth muscle cells mediated by PDGF," *Atherosclerosis*, vol. 84, no. 2-3, pp. 183–188, 1990.
- [52] A. Nomoto, S. Mutoh, H. Hagihara, and I. Yamaguchi, "Smooth muscle cell migration induced by inflammatory cell products and its inhibition by a potent calcium antagonist, nilvadipine," *Atherosclerosis*, vol. 72, no. 2-3, pp. 213–219, 1988.
- [53] H. Shimokawa, A. Ito, Y. Fukumoto et al., "Chronic treatment with interleukin-1 beta induces coronary intimal lesions and vasospastic responses in pigs in vivo. The role of platelet-derived growth factor," *The Journal of Clinical Investigation*, vol. 97, no. 3, pp. 769–776, 1996.
- [54] S. E. Francis, N. J. Camp, R. M. Dewberry et al., "Interleukin-1 receptor antagonist gene polymorphism and coronary artery disease," *Circulation*, vol. 99, no. 7, pp. 861–866, 1999.
- [55] I. Ikonomidis, J. P. Lekakis, M. Nikolaou et al., "Inhibition of interleukin-1 by anakinra improves vascular and left ventricular function in patients with rheumatoid arthritis," *Circulation*, vol. 117, no. 20, pp. 2662–2669, 2008.
- [56] S. Vallejo, E. Palacios, T. Romacho, L. Villalobos, C. Peiro, and C. F. Sanchez-Ferrer, "The interleukin-1 receptor antagonist anakinra improves endothelial dysfunction in streptozotocin-induced diabetic rats," *Cardiovascular Diabetology*, vol. 13, no. 1, p. 158, 2014.
- [57] P. M. Ridker, B. M. Everett, T. Thuren et al., "Antiinflammatory therapy with canakinumab for atherosclerotic disease," *The New England Journal of Medicine*, vol. 377, no. 12, pp. 1119–1131, 2017.
- [58] S. Wassmann, M. Stumpf, K. Strehlow et al., "Interleukin-6 induces oxidative stress and endothelial dysfunction by over-expression of the angiotensin II type 1 receptor," *Circulation Research*, vol. 94, no. 4, pp. 534–541, 2004.
- [59] E. Esteve, A. Castro, A. Lopez-Bermejo, J. Vendrell, W. Ricart, and J. M. Fernandez-Real, "Serum interleukin-6 correlates with endothelial dysfunction in healthy men independently of insulin sensitivity," *Diabetes Care*, vol. 30, no. 4, pp. 939–945, 2007.
- [60] M. Naya, T. Tsukamoto, K. Morita et al., "Plasma interleukin-6 and tumor necrosis factor-α can predict coronary endothelial dysfunction in hypertensive patients," *Hypertension Research*, vol. 30, no. 6, pp. 541–548, 2007.

- [61] H. Nawawi, N. S. Osman, R. Annuar, B. A. Khalid, and K. Yusoff, "Soluble intercellular adhesion molecule-1 and interleukin-6 levels reflect endothelial dysfunction in patients with primary hypercholesterolaemia treated with atorvastatin," *Atherosclerosis*, vol. 169, no. 2, pp. 283–291, 2003.
- [62] A. Mahmud and J. Feely, "Arterial stiffness is related to systemic inflammation in essential hypertension," *Hyperten*sion, vol. 46, no. 5, pp. 1118–1122, 2005.
- [63] S. Kaptoge, S. R. Seshasai, P. Gao et al., "Inflammatory cytokines and risk of coronary heart disease: new prospective study and updated meta-analysis," *European Heart Journal*, vol. 35, no. 9, pp. 578–589, 2014.
- [64] A. D. Protogerou, E. Zampeli, K. Fragiadaki, K. Stamatelopoulos, C. Papamichael, and P. P. Sfikakis, "A pilot study of endothelial dysfunction and aortic stiffness after interleukin-6 receptor inhibition in rheumatoid arthritis," *Atherosclerosis*, vol. 219, no. 2, pp. 734–736, 2011.
- [65] B. C. Bacchiega, A. B. Bacchiega, M. J. Usnayo, R. Bedirian, G. Singh, and G. D. Pinheiro, "Interleukin 6 inhibition and coronary artery disease in a high-risk population: a prospective community-based clinical study," *Journal of the American Heart Association*, vol. 6, no. 3, article e005038, 2017.
- [66] P. Ruiz-Limon, R. Ortega, I. Arias de la Rosa et al., "Tocilizumab improves the proatherothrombotic profile of rheumatoid arthritis patients modulating endothelial dysfunction, NETosis, and inflammation," *Translational Research*, vol. 183, pp. 87–103, 2017.
- [67] A. Floris, M. Piga, A. Cauli, and A. Mathieu, "Predictors of flares in systemic lupus erythematosus: preventive therapeutic intervention based on serial anti-dsDNA antibodies assessment. Analysis of a monocentric cohort and literature review," *Autoimmunity Reviews*, vol. 15, no. 7, pp. 656–663, 2016
- [68] J. Braun, "Methotrexate: optimizing the efficacy in rheumatoid arthritis," *Therapeutic Advances in Musculoskeletal Disease*, vol. 3, no. 3, pp. 151–158, 2011.
- [69] H. K. Choi, M. A. Hernan, J. D. Seeger, J. M. Robins, and F. Wolfe, "Methotrexate and mortality in patients with rheumatoid arthritis: a prospective study," *The Lancet*, vol. 359, no. 9313, pp. 1173–1177, 2002.
- [70] D. Krause, B. Schleusser, G. Herborn, and R. Rau, "Response to methotrexate treatment is associated with reduced mortality in patients with severe rheumatoid arthritis," *Arthritis & Rheumatism*, vol. 43, no. 1, pp. 14–21, 2000.
- [71] M. C. Wasko, A. Dasgupta, H. Hubert, J. F. Fries, and M. M. Ward, "Propensity-adjusted association of methotrexate with overall survival in rheumatoid arthritis," *Arthritis & Rheumatism*, vol. 65, no. 2, pp. 334–342, 2013.
- [72] B. Bannwarth, F. Pehourcq, T. Schaeverbeke, and J. Dehais, "Clinical pharmacokinetics of low-dose pulse methotrexate in rheumatoid arthritis," *Clinical Pharmacokinetics*, vol. 30, no. 3, pp. 194–210, 1996.
- [73] S. Pan, L. K. Stamp, S. B. Duffull et al., "Assessment of the relationship between methotrexate polyglutamates in red blood cells and clinical response in patients commencing methotrexate for rheumatoid arthritis," *Clinical Pharmacoki*netics, vol. 53, no. 12, pp. 1161–1170, 2014.
- [74] M. C. de Rotte, E. den Boer, P. H. de Jong et al., "Methotrexate polyglutamates in erythrocytes are associated with lower disease activity in patients with rheumatoid arthritis," *Annals* of the Rheumatic Diseases, vol. 74, no. 2, pp. 408–414, 2015.

[75] K. Inoue and H. Yuasa, "Molecular basis for pharmacokinetics and pharmacodynamics of methotrexate in rheumatoid arthritis therapy," *Drug Metabolism and Pharmacokinetics*, vol. 29, no. 1, pp. 12–19, 2014.

- [76] G. Hasko and B. Cronstein, "Regulation of inflammation by adenosine," *Frontiers in Immunology*, vol. 4, p. 85, 2013.
- [77] B. N. Cronstein, D. Naime, and E. Ostad, "The antiinflammatory mechanism of methotrexate. Increased adenosine release at inflamed sites diminishes leukocyte accumulation in an in vivo model of inflammation," *The Journal of Clinical Investigation*, vol. 92, no. 6, pp. 2675–2682, 1993.
- [78] D. G. Hardie, "AMP-activated protein kinase-an energy sensor that regulates all aspects of cell function," *Genes & Development*, vol. 25, no. 18, pp. 1895–1908, 2011.
- [79] L. Antonioli, R. Colucci, C. Pellegrini et al., "The AMPK enzyme-complex: from the regulation of cellular energy homeostasis to a possible new molecular target in the management of chronic inflammatory disorders," *Expert Opinion* on Therapeutic Targets, vol. 20, no. 2, pp. 179–191, 2016.
- [80] F. Boin, G. L. Erre, A. M. Posadino et al., "Oxidative stress-dependent activation of collagen synthesis is induced in human pulmonary smooth muscle cells by sera from patients with scleroderma-associated pulmonary hypertension," Orphanet Journal of Rare Diseases, vol. 9, no. 1, p. 123, 2014.
- [81] M. Igata, H. Motoshima, K. Tsuruzoe et al., "Adenosine monophosphate-activated protein kinase suppresses vascular smooth muscle cell proliferation through the inhibition of cell cycle progression," *Circulation Research*, vol. 97, no. 8, pp. 837–844, 2005.
- [82] Y. Ma, L. Li, Y. Shao, X. Bai, T. Bai, and X. Huang, "Methotrexate improves perivascular adipose tissue/endothelial dysfunction via activation of AMPK/eNOS pathway," *Molecular Medicine Reports*, vol. 15, no. 4, pp. 2353–2359, 2017.
- [83] R. Micha, F. Imamura, M. Wyler von Ballmoos et al., "Systematic review and meta-analysis of methotrexate use and risk of cardiovascular disease," *The American Journal of Cardiology*, vol. 108, no. 9, pp. 1362–1370, 2011.
- [84] C. Roubille, V. Richer, T. Starnino et al., "The effects of tumour necrosis factor inhibitors, methotrexate, nonsteroidal anti-inflammatory drugs and corticosteroids on cardiovascular events in rheumatoid arthritis, psoriasis and psoriatic arthritis: a systematic review and meta-analysis," *Annals of the Rheumatic Diseases*, vol. 74, no. 3, pp. 480–489, 2015.
- [85] M. Seitz, M. Zwicker, and P. Loetscher, "Effects of methotrexate on differentiation of monocytes and production of cytokine inhibitors by monocytes," *Arthritis & Rheumatism*, vol. 41, no. 11, pp. 2032–2038, 1998.
- [86] C. K. Edwards 3rd, A. M. Bendele, L. I. Reznikov et al., "Soluble human p55 and p75 tumor necrosis factor receptors reverse spontaneous arthritis in transgenic mice expressing transmembrane tumor necrosis factor α," *Arthritis & Rheumatism*, vol. 54, no. 9, pp. 2872–2885, 2006.
- [87] F. G. Sajjadi, K. Takabayashi, A. C. Foster, R. C. Domingo, and G. S. Firestein, "Inhibition of TNF-alpha expression by adenosine: role of A3 adenosine receptors," *Journal of Immunology*, vol. 156, no. 9, pp. 3435–3442, 1996.
- [88] A. Bulgarelli, A. A. Martins Dias, B. Caramelli, and R. C. Maranhao, "Treatment with methotrexate inhibits atherogenesis in cholesterol-fed rabbits," *Journal of Cardiovascular Pharmacology*, vol. 59, no. 4, pp. 308–314, 2012.

[89] C. C. Thornton, F. Al-Rashed, D. Calay et al., "Methotrexate-mediated activation of an AMPK-CREB-dependent pathway: a novel mechanism for vascular protection in chronic systemic inflammation," *Annals of the Rheumatic Diseases*, vol. 75, no. 2, pp. 439–448, 2016.

- [90] E. Yamasaki, Y. Soma, Y. Kawa, and M. Mizoguchi, "Methotrexate inhibits proliferation and regulation of the expression of intercellular adhesion molecule-1 and vascular cell adhesion molecule-1 by cultured human umbilical vein endothelial cells," *The British Journal of Dermatology*, vol. 149, no. 1, pp. 30–38, 2003.
- [91] T. J. Kalogeris, C. Baines, and R. J. Korthuis, "Adenosine prevents TNFα-induced decrease in endothelial mitochondrial mass via activation of eNOS-PGC-1α regulatory axis," PLoS One, vol. 9, no. 6, article e98459, 2014.
- [92] E. S. Cohen, W. R. Law, C. R. Easington et al., "Adenosine deaminase inhibition attenuates microvascular dysfunction and improves survival in sepsis," *American Journal of Respiratory and Critical Care Medicine*, vol. 166, no. 1, pp. 16–20, 2002.
- [93] C. M. Summers, A. L. Hammons, J. Arora et al., "Methotrexate modulates folate phenotype and inflammatory profile in EA.hy 926 cells," *European Journal of Pharmacology*, vol. 732, pp. 60–67, 2014.
- [94] J. Li, Y. Wang, Y. Wang et al., "Pharmacological activation of AMPK prevents Drp1-mediated mitochondrial fission and alleviates endoplasmic reticulum stress-associated endothelial dysfunction," *Journal of Molecular and Cellular Cardiol*ogy, vol. 86, pp. 62–74, 2015.
- [95] X. Hou and F. Pei, "Estradiol inhibits cytokine-induced expression of VCAM-1 and ICAM-1 in cultured human endothelial cells via AMPK/PPARα activation," Cell Biochemistry and Biophysics, vol. 72, no. 3, pp. 709–717, 2015
- [96] A. Quan, Y. Pan, K. K. Singh et al., "Cardiovascular inflammation is reduced with methotrexate in diabetes," *Molecular and Cellular Biochemistry*, vol. 432, no. 1-2, pp. 159–167, 2017
- [97] S. M. Hassanian, P. Dinarvand, and A. R. Rezaie, "Adenosine regulates the proinflammatory signaling function of thrombin in endothelial cells," *Journal of Cellular Physiology*, vol. 229, no. 9, pp. 1292–1300, 2014.
- [98] M. G. Bouma, F. A. van den Wildenberg, and W. A. Buurman, "Adenosine inhibits cytokine release and expression of adhesion molecules by activated human endothelial cells," *The American Journal of Physiology*, vol. 270, 2 Part 1, pp. C522–C529, 1996.
- [99] J. C. Shryock and L. Belardinelli, "Adenosine and adenosine receptors in the cardiovascular system: biochemistry, physiology, and pharmacology," *The American Journal of Cardiol*ogy, vol. 79, no. 12, Supplement 1, pp. 2–10, 1997.
- [100] R. A. Olsson and J. D. Pearson, "Cardiovascular purinoceptors," *Physiological Reviews*, vol. 70, no. 3, pp. 761–845, 1990.
- [101] L. Stella, V. de Novellis, I. Marabese et al., "The role of A3 adenosine receptors in central regulation of arterial blood pressure," *British Journal of Pharmacology*, vol. 125, no. 3, pp. 437–440, 1998.
- [102] M. Koupenova, H. Johnston-Cox, and K. Ravid, "Regulation of atherosclerosis and associated risk factors by adenosine and adenosine receptors," *Current Atherosclerosis Reports*, vol. 14, no. 5, pp. 460–468, 2012.

[103] K. Varani, F. Portaluppi, S. Merighi, E. Ongini, L. Belardinelli, and P. A. Borea, "Caffeine alters A_{2A} adenosine receptors and their function in human platelets," *Circulation*, vol. 99, no. 19, pp. 2499–2502, 1999.

- [104] L. Antonioli, C. Blandizzi, B. Csoka, P. Pacher, and G. Hasko, "Adenosine signalling in diabetes mellitus-pathophysiology and therapeutic considerations," *Nature Reviews. Endocrinol*ogy, vol. 11, no. 4, pp. 228–241, 2015.
- [105] D. Li, D. Wang, Y. Wang, W. Ling, X. Feng, and M. Xia, "Adenosine monophosphate-activated protein kinase induces cholesterol efflux from macrophage-derived foam cells and alleviates atherosclerosis in apolipoprotein Edeficient mice," *The Journal of Biological Chemistry*, vol. 285, no. 43, pp. 33499–33509, 2010.
- [106] Z. Chen, I. C. Peng, W. Sun et al., "AMP-activated protein kinase functionally phosphorylates endothelial nitric oxide synthase Ser633," *Circulation Research*, vol. 104, no. 4, pp. 496–505, 2009.
- [107] F. Goirand, M. Solar, Y. Athea et al., "Activation of AMP kinase α1 subunit induces aortic vasorelaxation in mice," *The Journal of Physiology*, vol. 581, no. 3, pp. 1163–1171, 2007.
- [108] D. Nagata, R. Takeda, M. Sata et al., "AMP-activated protein kinase inhibits angiotensin II-stimulated vascular smooth muscle cell proliferation," *Circulation*, vol. 110, no. 4, pp. 444–451, 2004.
- [109] R. J. Ford, S. R. Teschke, E. B. Reid, K. K. Durham, J. T. Kroetsch, and J. W. Rush, "AMP-activated protein kinase activator AICAR acutely lowers blood pressure and relaxes isolated resistance arteries of hypertensive rats," *Journal of Hypertension*, vol. 30, no. 4, pp. 725–733, 2012.
- [110] E. S. Buhl, N. Jessen, R. Pold et al., "Long-term AICAR administration reduces metabolic disturbances and lowers blood pressure in rats displaying features of the insulin resistance syndrome," *Diabetes*, vol. 51, no. 7, pp. 2199–2206, 2002
- [111] N. Wu, B. Zheng, A. Shaywitz et al., "AMPK-dependent degradation of TXNIP upon energy stress leads to enhanced glucose uptake via GLUT1," *Molecular Cell*, vol. 49, no. 6, pp. 1167–1175, 2013.
- [112] S. L. McGee, B. J. van Denderen, K. F. Howlett et al., "AMP-activated protein kinase regulates GLUT4 transcription by phosphorylating histone deacetylase 5," *Diabetes*, vol. 57, no. 4, pp. 860–867, 2008.
- [113] A. S. Marsin, L. Bertrand, M. H. Rider et al., "Phosphorylation and activation of heart PFK-2 by AMPK has a role in the stimulation of glycolysis during ischaemia," *Current Biology*, vol. 10, no. 20, pp. 1247–1255, 2000.
- [114] A. S. Marsin, C. Bouzin, L. Bertrand, and L. Hue, "The stimulation of glycolysis by hypoxia in activated monocytes is mediated by AMP-activated protein kinase and inducible 6-phosphofructo-2-kinase," *The Journal of Biological Chemistry*, vol. 277, no. 34, pp. 30778–30783, 2002.
- [115] Z. Xie, J. Zhang, J. Wu, B. Viollet, and M. H. Zou, "Upregulation of mitochondrial uncoupling protein-2 by the AMP-activated protein kinase in endothelial cells attenuates oxidative stress in diabetes," *Diabetes*, vol. 57, no. 12, pp. 3222–3230, 2008.
- [116] X. N. Li, J. Song, L. Zhang et al., "Activation of the AMPK-FOXO3 pathway reduces fatty acid-induced increase in intracellular reactive oxygen species by upregulating thioredoxin," *Diabetes*, vol. 58, no. 10, pp. 2246–2257, 2009.

[117] S. Hirata, T. Matsubara, R. Saura, H. Tateishi, and K. Hirohata, "Inhibition of in vitro vascular endothelial cell proliferation and in vivo neovascularization by low-dose methotrexate," *Arthritis & Rheumatism*, vol. 32, no. 9, pp. 1065–1073, 1989.

- [118] T. Annussek, T. Szuwart, J. Kleinheinz, C. Koiky, and K. Wermker, "In vitro inhibition of HUVECs by low dose methotrexate insights into oral adverse events," *Head & Face Medicine*, vol. 10, no. 1, p. 19, 2014.
- [119] L. Gao, K. Chalupsky, E. Stefani, and H. Cai, "Mechanistic insights into folic acid-dependent vascular protection: dihydrofolate reductase (DHFR)-mediated reduction in oxidant stress in endothelial cells and angiotensin II-infused mice: a novel HPLC-based fluorescent assay for DHFR activity," *Journal of Molecular and Cellular Cardiology*, vol. 47, no. 6, pp. 752–760, 2009.
- [120] J. L. Pastrana, X. Sha, A. Virtue et al., "Regulatory T cells and atherosclerosis," *Journal of Clinical & Experimental Cardiol*ogy, vol. 1, Supplement 12, p. 2, 2013.
- [121] B. M. Everett, A. D. Pradhan, D. H. Solomon et al., "Rationale and design of the cardiovascular inflammation reduction trial: a test of the inflammatory hypothesis of atherothrombosis," *American Heart Journal*, vol. 166, no. 2, pp. 199–207.e15, 2013.

















Submit your manuscripts at https://www.hindawi.com











Behavioural Neurology











